

EPPO Datasheet: '*Candidatus Liberibacter asiaticus*'

Last updated: 2020-06-09

This datasheet covers the three bacterial species that are associated with huanglongbing (or citrus greening). Huanglongbing is transmitted by two psyllid vectors ([Diaphorina citri](#) and [Trioza erytreae](#)) which are covered in two other separate datasheets.

IDENTITY

Preferred name: '*Candidatus Liberibacter asiaticus*'

Authority: Jagoueix, Bové & Garnier

Taxonomic position: Bacteria: Proteobacteria: Alphaproteobacteria: Rhizobiales: Phyllobacteriaceae

Other scientific names: *Citrus greening bacterium* (heat-tolerant strain), *Liberibacter asiaticum*, *Liberibacter asiaticus*

Common names: blotchy mottle disease of citrus, citrus dieback, decline of citrus, greening of citrus, huanglongbing, leaf mottling of citrus, likubin, vein phloem degeneration of citrus deactivated, yellow branch of citrus, yellow shoot disease, yellow shoot of citrus

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EPPO Categorization: A1 list

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EU Categorization: Quarantine pest ((EU) 2019/2072 Annex II A)

EPPO Code: LIBEAS



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Notes on taxonomy and nomenclature

The disease name 'greening' was the first English name adopted by the scientific literature probably because of the influence of South Africa research, as it was the disease name in that country. In 1995 the International Organization of Citrus Virologists (IOCV) decided to adopt the original Chinese name of 'huanglongbing' as official (Gottwald, 2010). The acronym HLB is also widely used in the literature.

For many years, abiotic factors such as mineral deficiencies (or toxicities) and water logging were thought to be the causes of huanglongbing. In the 1960s, it was suspected that the causal agent was a virus or a mycoplasma-like organism (MLO). The discovery by transmission electron microscopy of cell-walled bacteria in affected plants demonstrated that true bacteria were present (Laflèche & Bové, 1970; Gottwald *et al.*, 2007).

Three taxonomic entities are currently associated with huanglongbing symptoms. They are all fastidious, phloem-limited, Gram-negative bacteria with a peptidoglycan-containing cell wall (Moll and Martin, 1973; Garnier *et al.*, 1984). Molecular and phylogenetic analyses have demonstrated that they belong to the family Phyllobacteriaceae. The first proposed names were '*Candidatus Liberobacter africanum*' and '*Candidatus Liberobacter asiaticum*' Jagoueix, Bové & Garnier (Jagoueix *et al.*, 1994). They were then changed to '*Candidatus Liberibacter africanus*' and '*Candidatus Liberibacter asiaticus*' to follow the rules of the International Code of Nomenclature of Bacteria (Garnier *et al.*, 2000). In 2005, a third species was found in Brazil and called '*Ca. Liberibacter americanus*' (Teixeira *et al.*, 2005 b). As these bacteria have not yet been cultivated in axenic culture, the Koch's postulates have not been fulfilled to confirm that they are the causal agents of the disease. Consequently, according to the rules of taxonomy they must be named '*Candidatus*,' an interim taxonomic status. In the past, two forms of the disease were reported: a heat-tolerant 'Asian form' now identified as '*Ca. Liberibacter asiaticus*', and a heat-sensitive 'African form' now identified as '*Ca. Liberibacter africanus*'. '*Ca. Liberibacter americanus*' has also been shown to be heat sensitive (Lopes *et al.*, 2009 b).

Several subspecies have also been proposed. In 1995 a new strain was detected in *Calodendrum capense* from South Africa and named '*Candidatus Liberibacter africanus* subsp. *capensis*' (Garnier *et al.*, 2000). More recently, four new subspecies have been proposed: '*Candidatus Liberibacter africanus* subsp. *clausenae*', '*Candidatus Liberibacter*

africanus subsp. vepridis', 'Candidatus Liberibacter africanus subsp. zanthoxyli' and 'Candidatus Liberibacter africanus subsp. teclae' (Roberts *et al.*, 2015; Roberts & Pietersen, 2017) but they are not considered as validly published.

HOSTS

The three species of 'Ca. Liberibacter' infect species of *Citrus* and other genera within the Rutaceae family. There is no available information about differences in host range between 'Ca. Liberibacter asiaticus' and 'Ca. Liberibacter africanus', and consequently the host list is the same for these two species (ANSES, 2019; EFSA, 2019). Both can multiply and colonize many *Citrus* spp., but the most severe symptoms are found on sweet orange (*C. sinensis*), mandarin (*C. reticulata*) and tangelo (*C. reticulata* x *C. paradisi*). Somewhat less severe symptoms are found on lemon (*C. limon*), grapefruit (*C. paradisi*), Rangpur lime (*C. limonia*), Palestinian sweet lime (*C. limettioides*), rough lemon (*C. jambhiri*), kumquat (*Fortunella* spp.) and citron (*C. medica*) (McClellan & Schwarz, 1970). Symptoms are even weaker on lime (*C. aurantiifolia*) and pummelo (*C. grandis*).

Some citrus-related plants have been confirmed as hosts for the disease, namely *Severinia buxifolia*, *Limonia acidissima* and *Vepris lanceolata* (ANSES, 2019). The ornamental Rutaceae, *Murraya paniculata*, is a very important host for 'Ca. Liberibacter asiaticus' in American countries and *M. koenigii* (*Berberis koenigii*) is also recorded as a host in other countries. There is some confusion concerning the taxonomic distinction between *M. paniculata* and *Murraya exotica*, the latter being more susceptible to bacterial infection and more attractive to the vector *Diaphorina citri*.

Other species of Rutaceae have been infected by experimental inoculation, but apparently there are no records of natural infections. Both bacterial species have been experimentally transmitted by *Cuscuta campestris*, from citrus to the following non-rutaceous hosts: *Catharanthus roseus*, *Nicotiana glauca*, *N. tabacum* and *Solanum lycopersicum* (Garnier and Bové, 1983; ANSES, 2019; EFSA, 2019).

For 'Ca. Liberibacter americanus', data are scarce, and it has only been reported on sweet orange (*C. sinensis*), mandarin (*Citrus reticulata*), tangor (*C. reticulata* x *C. sinensis*) and *M. paniculata* (Bové 2006; Lopes *et al.* 2010).

In summary, genera of Rutaceae with species affected by huanglongbing are: *Atalantia*, *Balsamocitrus*, *Calodendron*, *Citroncirus*, *Citroncirus* x (*Citrange*), *Citrofortunella*, *Citrus*, *Citrus* x *Limonia*, *Citrus* x *Tangelo*, *Clausena*, *Fortunella*, *Limonia*, *Murraya*, *Poncirus*, *Severinia*, *Swinglea*, *Toddalia* and *Vepris*. In addition, weeds of non Rutaceae plants such as some in the genera *Cleome*, *Pisonia*, *Pithecellobium*, *Trichostigma* and *Triphasia* may also be considered as hosts, since in Jamaica or China, species of these genera have been found infected in huanglongbing affected orchards (ANSES, 2019; EFSA, 2019 b).

Host list: *Atalantia buxifolia*, *Balsamocitrus dawei*, *Citroncirus webberi*, *Citroncirus*, *Citrus depressa*, *Citrus hystrix*, *Citrus limonimeditica*, *Citrus macroptera*, *Citrus maxima*, *Citrus medica*, *Citrus reticulata*, *Citrus trifoliata*, *Citrus* x *amblycarpa*, *Citrus* x *aurantiifolia*, *Citrus* x *aurantium* var. *paradisi*, *Citrus* x *aurantium* var. *sinensis*, *Citrus* x *aurantium*, *Citrus* x *junos*, *Citrus* x *limon* var. *limettioides*, *Citrus* x *limon*, *Citrus* x *limonia* var. *jambhiri*, *Citrus* x *limonia* var. *volkammeriana*, *Citrus* x *limonia*, *Citrus* x *nobilis*, *Citrus* x *tangelo*, *Citrus*, *Clausena indica*, *Clausena lansium*, *Cleome rutidosperma*, *Fortunella margarita*, *Fortunella*, *Limonia acidissima*, *Murraya paniculata*, *Murraya sumatrana*, *Pisonia aculeata*, *Rutaceae*, *Swinglea glutinosa*, *Toddalia*, *Trichostigma octandrum*, x *Citrofortunella microcarpa*

GEOGRAPHICAL DISTRIBUTION

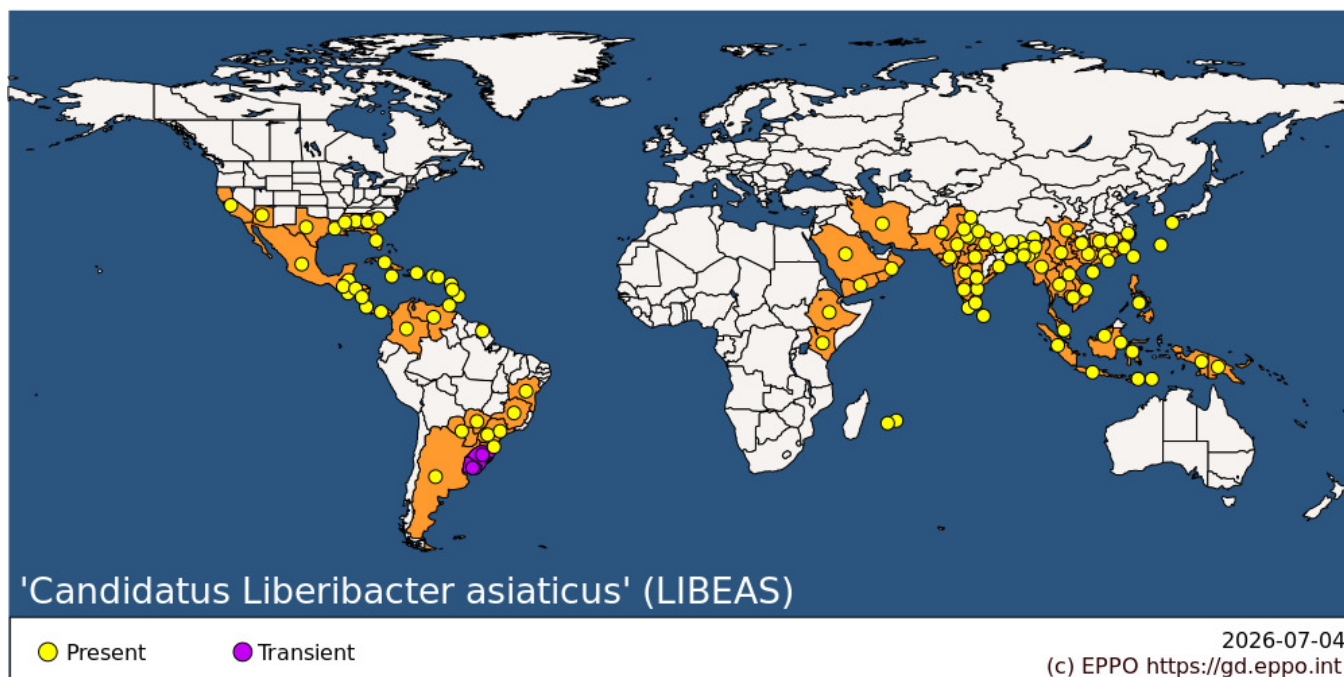
Huanglongbing was probably first observed in Asia in the 18th century when a severe disease of unknown origin called 'citrus dieback' was recorded in the central provinces of India. Then, in 1919, a disease causing yellowing and leaf mottle symptoms was reported in Southern China as present since the late 19th century and farmers called it 'huanglongbing' that means yellow shoot disease. Similar symptoms were reported in Pakistan in 1927 and in Southern China in 1943 (Gottwald *et al.*, 2007). From the 1920s, this new citrus disease was also described in several other Asian countries (Philippines, Taiwan, Indonesia), as well as in other areas of China. During the same period, in 1928 similar disorders were reported in South Africa as 'yellow branch disease', later called 'greening', and afterwards reported in other African countries. For decades, huanglongbing (or greening) has been considered

limited to Asian and African countries, but in 2004 it was found in Brazil and in 2005 in Florida (USA). In the following years, it was also found in other USA states and many American countries (Bové, 2006; Dala-Paula *et al.*, 2019).

The available information about the geographical distribution does not always indicate if the species identified in each country was asiaticus or africanus, especially in the old literature. The Mediterranean area and most of the Middle East, Australia, New Zealand, New Caledonia and small Pacific islands are still free from the disease (Duran-Vila *et al.*, 2014; Siverio *et al.*, 2017).

Regarding disease vectors, *Diaphorina citri* has recently been reported in the EPPO region in Cyprus and Israel (Melifronidou?Pantelidou *et al.*, 2025), and *Trioza erytreae* is present with restricted distribution in Spain, (including Canary Islands), and in Portugal, (including Madeira Island) (Pérez-Otero *et al.*, 2015; Siverio *et al.*, 2017; Arenas-Arenas *et al.*, 2018).

The map below shows the world distribution of '*Ca. Liberibacter asiaticus*'. Click on the links to view the distributions of '*Ca. Liberibacter africanus*' and '*Ca. Liberibacter americanus*'.



Africa: Ethiopia, Kenya, Mauritius, Reunion

Asia: Bangladesh, Bhutan, Cambodia, China (Fujian, Guangdong, Guangxi, Guizhou, Hainan, Hunan, Jiangxi, Sichuan, Xianggang (Hong Kong), Yunnan, Zhejiang), East Timor, India (Andhra Pradesh, Arunachal Pradesh, Assam, Bihar, Delhi, Gujarat, Haryana, Himachal Pradesh, Jammu & Kashmir, Karnataka, Kerala, Madhya Pradesh, Maharashtra, Manipur, Meghalaya, Mizoram, Nagaland, Odisha, Punjab, Rajasthan, Sikkim, Tamil Nadu, Telangana, Tripura, Uttarakhand, Uttar Pradesh, West Bengal), Indonesia (Irian Jaya, Java, Kalimantan, Nusa Tenggara, Sulawesi, Sumatra), Iran, Islamic Republic of, Japan (Kyushu, Ryukyu Archipelago), Lao People's Democratic Republic, Malaysia (Sarawak, West), Myanmar, Nepal, Oman, Pakistan, Philippines, Saudi Arabia, Sri Lanka, Taiwan, Thailand, Vietnam, Yemen

North America: Mexico, United States of America (Alabama, Arizona, California, Florida, Georgia, Louisiana, Mississippi, South Carolina, Texas)

Central America and Caribbean: Barbados, Belize, Costa Rica, Cuba, Dominica, Dominican Republic, El Salvador, Guadeloupe, Guatemala, Honduras, Jamaica, Martinique, Nicaragua, Panama, Puerto Rico, Trinidad and Tobago, Virgin Islands (US)

South America: Argentina, Brazil (Bahia, Mato Grosso do Sul, Minas Gerais, Parana, Rio Grande do Sul, Santa Catarina, Sao Paulo), Colombia, French Guiana, Paraguay, Uruguay, Venezuela

Oceania: Papua New Guinea

BIOLOGY

Location within the plant

The three '*Ca. Liberibacter*' reside within phloem tissues, being restricted to the inside of sieve tubes (Folimonova and Achor, 2010), although '*Ca. Liberibacter asiaticus*' has been reported in companion cells (Fu *et al.*, 2015). Systemic infection of the host plant follows the direction of phloem sap translocation, moving in a passive way from leaves to roots, flushes, and fruits. The movement of '*Ca. Liberibacter*' occurs primarily in a vertical direction through the sieve pores, rather than horizontally to adjacent sieve tubes (Wang *et al.*, 2017). Bacterial infection causes accumulated starch in the sieve elements, ultrastructural changes of phloem tissue, plugged sieve pores, and eventually phloem disruption (da Graça *et al.*, 2016). The bacteria multiply very well within the roots suggesting that early invasion of roots by these bacteria leads to root decline before the appearance of foliar symptoms (Johnson *et al.*, 2014).

Transmission

In 1956, Lin described the symptoms of the disease in China and in 1963 demonstrated the graft transmissibility of the disease (Zheng *et al.*, 2018). Later, this was confirmed in South Africa, as well as the transmission by the African citrus psyllid, *T. erythrae* (McClellan & Oberholzer, 1965). Almost at the same time, experiments in India and the Philippines demonstrated that another psyllid, *D. citri* (Capoor *et al.*, 1967) was also a vector of the disease in Asia. More recently, *Cacopsylla citrisuga* (Cen *et al.*, 2012) and *Diaphorina communis* (Donovan *et al.*, 2012), have also been reported as potential vectors of the disease. The two main psyllid vectors feed from the phloem sap of infected hosts and acquire the bacteria predominately from young shoots (EFSA, 2019 b). Then, they are disseminated into the plant, but with a heterogeneous distribution among the different organs. Once acquired, the bacteria remain in the hemolymph and the psyllid retains the ability to transmit the bacteria in a persistent manner throughout its lifespan.

Tolerance to temperature

The three *Liberibacter* species present differences in their tolerance to temperature and transmission by psyllid vectors. '*Ca. Liberibacter africanus*', transmitted by *T. erythrae* is heat-sensitive and disease symptoms do not develop in climates where temperatures above 30°C are reached several hours a day (Bové *et al.*, 1974). '*Ca. Liberibacter asiaticus*' which is present in Asia and America, is heat-tolerant and withstands high temperatures, its optimum ranging from 24 to 32°C and is transmitted by *D. citri*. Experimentally and also naturally in some countries *T. erythrae* and *D. citri* can also transmit the Asian and African forms, respectively (Massonié *et al.*, 1976; Lallemand *et al.*, 1986; Ajene *et al.*, 2020).

'*Ca. Liberibacter americanus*' was detected in Brazil with *D. citri* as its vector and it can also be transmitted by graft inoculations under greenhouse conditions (Teixeira *et al.*, 2005 b). '*Ca. Liberibacter americanus*' is heat sensitive and was discovered in central and southern Brazilian regions, where the annual number of hours above 30°C is two to five times lower than that in the extreme northern and western regions. In experimental conditions, trees inoculated with '*Ca. Liberibacter asiaticus*' had high bacterial titres and showed symptoms at 32 and 35°C, but not at 38°C, while temperatures of 32°C or above were detrimental to '*Ca. Liberibacter americanus*' (Lopes *et al.*, 2009 a and b). Mixed infections of two of these species have been also reported (Coletta-Filho *et al.*, 2005).

Epidemics

Epidemics of huanglongbing are established by introduction of infected plant material followed by natural transmission due to vectors. The unintentional entry of infected plant material establishes the disease in new areas or countries and subsequent unregulated movement of plants can have disastrous consequences. Natural transmission appears to be related to high vector populations and the extensiveness of the inoculum reservoirs. As psyllid migrations appear to be highest when host plants are flushing, natural spread is probably greatest in late spring and perhaps in other periods when new citrus flushes are available and psyllid populations are high (Gottwald *et al.*, 2007; Gottwald, 2010).

At field level, in areas where huanglongbing is present, aggregations of infected trees can be quite large, for example encompassing more than 1 600 trees in Florida (Gottwald *et al.*, 2007). This does not mean that every tree in these

areas will become infected, but that a high proportion of them will be so. Vectors apparently spread the pathogen to either adjacent or nearby trees only a few spaces away. The secondary foci are at variable distance from the main cluster of disease and when vectors move, (naturally for feeding or when disturbed by cultural practices), they occasionally move at least 25 to 50 m (Gottwald, 2010). However, as most of the spatial and temporal data analysed up to now were collected by visual assessments, it is probable that more accurate detection methods will improve the understanding of the disease epidemiology. By using PCR tests, it has been demonstrated that visual assessment largely underestimated disease prevalence. The number of trees found to be infected using PCR tests was more than double the number of positive results from visual assessment (Gottwald, 2010).

The spatial and temporal dynamics of the bacterial species associated with huanglongbing in citrus orchards have been investigated in different parts of the world. In South Africa, epidemics of '*Ca. Liberibacter africanus*' have been frequently observed in young orchards, in areas where the disease is endemic and where numerous sources of inoculum are present. Studies have shown that in these areas, damage appeared more rapidly in young plants than in older ones, even when insecticide sprays were applied against *T. erytrae*. For '*Ca. Liberibacter asiaticus*' epidemics, when inoculum pressure and vector levels were high, disease prevalence reached almost 100 % in orchards managed without insecticides within three years after planting. In orchards treated with fenobucarb and imidacloprid, disease prevalence reached more than 70 % and 20 %, respectively. Similarly, in North and South American countries, the prevalence of symptomatic trees in new citrus plantations, surrounded by older and heavily affected blocks, reached 20% two years after plantation and 70% within five years (Gottwald, 2010).

Related to the co-habitation of several bacterial species in the same area, soon after the discovery of '*Ca. Liberibacter americanus*' in São Paulo, Brazil in 2004 (Coletta-Filho *et al.*, 2004), '*Ca. Liberibacter asiaticus*' was detected in a small number of samples and '*Ca. Liberibacter americanus*' in 95 % of symptomatic trees tested (Teixeira *et al.*, 2005 b). However, four years later, the situation had reversed, and most symptomatic trees when tested were found to be infected only with '*Ca. L. asiaticus*' (Lopes *et al.*, 2009 b).

Reviews of huanglongbing have been provided by McClean & Schwarz (1970), da Graça (1991), Da Graça & Korsten (2004), Bové (2006), Gottwald *et al.* (2007), Gottwald (2010), da Graça *et al.* (2016), Zheng *et al.* (2018), Li *et al.* (2020) among others.

DETECTION AND IDENTIFICATION

Symptoms

On infected plants, symptoms are expressed after a variable period of time after infection (from one to three years) which depends on several factors (e.g. initial bacterial inoculum, time of infection, environmental conditions, tree age, species/cultivar, sanitary status of the tree). Symptoms generally appear faster in young trees (Gottwald *et al.*, 2007). The general aspect of citrus trees affected by huanglongbing is open growth, stunting, twig dieback, sparse yellow foliage, and severe fruit drop. Certain symptoms are described as more frequently observed in some countries, such as in China, where leaf symptoms were seen initially on one limb of the tree causing yellow branch; or in South Africa where the disease is currently called greening because of the poorly coloured fruits and the inversion of colouration when maturing. Symptoms develop relatively slowly, and infected trees gradually decline in vigour and yield, and remain stunted or eventually die. The disease develops irregularly so that individual trees may show a mixture of normal and diseased sectors (Bové, 2006; Gottwald *et al.*, 2007; CABI, 2018; EFSA, 2019b). Symptoms are generally the same for the three '*Ca. Liberibacter*' species, although the Asian form is considered associated to more severe symptoms, because dieback can be more extensive and eventually resulting in tree death.

Phytoplasmas of several groups (16SrI, 16SrII, 16SrIII, 16SrVI and 16SrIX) have been reported associated with huanglongbing symptoms in Brazil, China, India, and Mexico (Wulff *et al.*, 2019). Zinc and copper deficiency may also cause symptoms similar to those described below.

On fruits

On infected plants, some fruits are under-developed and sometimes poorly coloured (greening). They often fail to develop normal fruit colour because at the time when the fruit changes from green to orange, affected fruit show colour inversion: the peduncular end of the fruit turns orange, while the styler end is still green, whereas on normal

fruit the coloration starts first at the stylar end. There is early fruit drop from affected branches reducing fruit harvest. Fruits from affected plants are smaller, lighter, and more acidic. They also have a bitter and salty taste and the juice quality is severely affected, making the fruits not exploitable for the industry. Inside, the columella is curved causing the fruit to be distorted and lopsided. Seeds in the affected fruit are usually abortive.

On leaves

Symptoms usually first appear as leaf yellowing followed by mottling and chlorosis in one shoot or sector of the tree. Later, leaf symptoms resemble nutritional deficiencies (zinc, copper, or manganese) but may vary depending on the bacterial strain. The larger leaves on the base of branches turn yellow along the main and secondary veins and later change to a blotchy-mottle. As the discoloration spreads away from the veins, the leaves become pale to light yellow with unevenly distributed dark green areas. Leaves on weak terminal twigs are small, up-right and show a variety of chlorotic patterns. This is the most characteristic foliar symptom, because the two halves of the leaf patterns of yellow and green areas are asymmetric, in contrast with the nutrient deficiencies, that are symmetric. Mature leaves often show irregular patches between the main veins. The veins are often prominent and yellow. Frequently, there is abundant leaf drop.

On trunk, limbs, and shoots

Twig dieback is abundant in chronically infected trees, but no symptoms are apparent on trunk. Histological symptoms are localized zones of necrotic phloem scattered through the vascular system of the leaf. Massive accumulation of starch in the plastids is observed together with aberrations in cambial activity and excessive phloem formation.

Morphology

The huanglongbing associated bacteria are variable in morphology but mainly are elongated sinuous rod-like structures, around 0.1-0.2 μm in diameter and around 1-2 μm long, but round forms of larger diameter can also be found (CABI, 2018). The bacterial cells can be observed by electron microscopy in the phloem of infected trees and in both vectors showing in thin-sections a characteristic double-membrane cell envelope (Garnier *et al.*, 1984).

Sequences of the three associated species have been published and they have a small genome that range from 1.1 to 1.2 Mb with a low GC content below 37 %. The average nucleotide identity (ANI) values between different strains of the same species are above 99 % and below 81% among the different species. Although genomic sequences of the three species are quite different the effect on the plants are similar, even if '*Ca. Liberibacter asiaticus*' is generally causing more severe symptoms. Complete type I secretion systems (T1SS) and one of its putative substrates such as serralyisin have been identified in '*Ca. Liberibacter asiaticus*' and '*Ca. Liberibacter africanus*' but not in '*Ca. Liberibacter americanus*'. A complete general secretory pathway (Sec) is present in the three liberibacters but significant differences among them have been identified in its putative substrates. Moreover, differences on the genes involved in lipopolysaccharides (LPSs) of the three liberibacters that might affect their induction of host plant defence have been also identified. As well, '*Ca. Liberibacter asiaticus*' and '*Ca. Liberibacter africanus*' show differences in flagella regulators as compared to '*Ca. Liberibacter americanus*', that maybe be also connected to a different induction of plant defences (Wang *et al.*, 2017).

Detection and inspection methods

Detailed protocols for surveillance, sampling and detection are indicated in the EPPO Standard PM 9/27 (2020).

Visual inspection

Visual symptoms are important for diagnosis in symptomatic plants and visual inspection is a routine method for huanglongbing eradication in countries where the disease is present, as well as for its surveillance in countries or areas where it is not present. Surveys must be carried out carefully, all trees in an orchard should be examined one by one, and a few minutes must be spent at each tree. The scouts in charge of the surveys should work in pairs, so that each tree is examined by the two scouts, one on each side of the row. In the case of orchards with adult trees, it is essential to examine the top of the trees. For this reason, in Brazil, high towers have been built onto tractors to permit

efficient observation of treetops (Bové, 2006). Finally, once affected trees have been identified, they should be removed as quickly as possible.

Yellow shoot and blotchy mottle on leaves are considered the most typical symptoms and can be used in field surveys as part of an initial diagnosis. However, some symptoms can be confused with nutritional disorders, deficiencies or with other diseases, because *Citrus tristeza virus*, stubborn, citrus blight, Australian citrus dieback, *Phytophthora spp.*, waterlogging or the use of marcots can produce similar blotchy mottle patterns, according to CABI (2018). A pest survey card on huanglongbing was prepared in the context of the EFSA mandate on plant pest surveillance, upon request by the European Commission to assist the Member States in planning annual survey activities (EFSA, 2019 a).

Symptoms of the infected trees in the aerial part are not always easy to distinguish from those due to other citrus diseases or abiotic factors. Yellow shoots, leaf blotchy mottle and lopsided fruits with colour inversion and aborted seeds, are quite specific but they do not always appear together on the same tree, and they can be distorted or masked by symptoms of other origins. In addition, trees can be latently infected for some months and the symptoms can appear even one or more years after infection (Lee *et al.*, 2015). Consequently, visual inspections can lead to false positives and negatives and complementary diagnosis (for symptomatic plants) or detection methods (for asymptomatic plants), in the laboratory and/or greenhouse must be performed.

Detection in plants

Different tools have been developed over time for the detection and/or identification of huanglongbing associated agents and are described with details and recommendations in the EPPO Standard PM 7/121 (EPPO, 2021). However, the low bacterial concentration in host plants and their uneven distribution may render their detection difficult (Gottwald, 2010). In the leaves the detection maybe problematic due to the spatial and seasonal patterns of pathogen movement in the plant (Wang *et al.*, 2017). Another difficulty is also that the bacteria associated with huanglongbing have not been cultured yet. Several reports claiming successful culture can be found in the literature, but there is still no experimental evidence to demonstrate that the described cultured organisms were really the causal agent of huanglongbing. However, studies indicating that the bacterium can be maintained in a biofilm form (but not yet in axenic culture) could be considered as a first step towards real isolation (Ha *et al.*, 2019).

For many years, electron microscopy was used as a diagnostic method, and is still useful to confirm the presence of the characteristic bacteria in the sieve tubes of trees presenting suspicious symptoms. In the late 1980s an enzyme-linked immunosorbent assay (ELISA) and an immunofluorescence test were developed. The presence of a specific fluorescent marker, gentisoyl glucoside in infected tissue was also used for detection. Biological indexing was employed for testing, as suspect material may be grafted onto sensitive indicator plants. Preferred indicator plants are Orlando tangelo and sweet orange seedlings.

More recently, several molecular methods have been developed for the detection and identification of the bacteria associated with huanglongbing. Conventional PCR is still used in some laboratories as screening test and for confirmation purposes for symptomatic material. For detection of '*Ca. Liberibacter africanus*' and '*Ca. Liberibacter asiaticus*' by conventional PCR, two sets of primers can be used (Jagoueix *et al.*, 1996 and Hocquellet *et al.*, 1999) and for '*Ca. Liberibacter americanus*' another set has been developed (Teixeira *et al.*, 2005 a). Conventional PCR can be also used for doubtful samples or for the first description of the disease in a new area.

However, real-time PCR is currently the preferred method for detecting these pathogens because of its high sensitivity and lower risk of contamination. Bertolini *et al.* (2014) developed a tissue-print (for plants) or squash (for vectors) methodology for performing a direct real-time PCR without the need of previous DNA extraction for detecting any '*Candidatus Liberibacter*' from symptomatic samples or suspected plants as well as for vectors in surveys; the samples can be directly imprinted in the field and sent by conventional mail to a laboratory to be processed (the imprints are non-infective samples). It is useful as a first screening and a good alternative for being used in the current situation of the EPPO countries in which the disease is still absent. The positive detections should be followed by specific real-time or conventional PCRs for the three huanglongbing associated bacteria and sequencing of the amplicons, to avoid false positives. Such method is well adapted to the countries where any of these bacteria has been detected and when the most important criterium is to avoid false negatives. It is simple, safe, and sensitive enough to be used for processing large numbers of symptomatic or suspected plants in surveys.

The real-time PCR based test using the TaqMan probe described by Li *et al.* (2006), is also very sensitive and it shows acceptable exclusivity and inclusivity criteria in the detection of these bacteria and Fu *et al.* (2019) also used the tissue printing system coupled with such real-time PCR with excellent results. For species identification, the primers and TaqMan probe described by Li *et al.* (2006); Morgan *et al.* (2012); Carlos *et al.* (2006); Teixeira *et al.* (2008) are also useful but all these protocols require a previous DNA extraction of the sample.

Detection in vectors

The different ‘*Ca. Liberibacter*’ associated to huanglongbing can also be detected in their psyllid vectors. Bertolini *et al.* (2014) developed a squash assay for individual psyllids followed by real-time PCR for *T. erytrae* and *D. citri*. Such methodology has been used in surveys conducted in Spain by Siverio *et al.* (2017). In the USA, ‘*Ca. Liberibacter asiaticus*’ has been found in *D. citri* several months or even years before symptoms appeared on infected plants (Manjunath *et al.*, 2008). In California (USA), the detection of the bacteria in *D. citri* was successfully used during the first surveys for the disease (Kumagai *et al.*, 2013). Testing psyllids has also proven valuable in assessing the status of plants for sale. Positive psyllids were found on average 9 months prior to the discovery of positive testing symptomatic plants in retail venues (Halbert *et al.*, 2012).

PATHWAYS FOR MOVEMENT

As the bacteria associated to huanglongbing are limited to the plant phloem, movement of infected host plant material (seedlings, plants, grafts, and rootstocks) are the main pathways for entry and spread over short and long distances (primary introduction). It is also considered that cut flowers, branches, foliage of host plants (in particular, *Murraya paniculata* and *Citrus hystrix*) can also be a pathway (ANSES, 2019). No transmission through seeds or fruits has been demonstrated yet. Hartung *et al.*, (2010) and Hilf (2011) found no evidence of seed transmission in hundreds of tested seedlings from seed collected from symptomatic fruit, although PCR tests on the fruit and seeds of infected plants (from which the seeds were used to produce the seedlings) were positive.

The psyllid vectors, *T. erytrae* and *D. citri* are responsible for disease spread over short and long distances (secondary spread), because they can both spread the disease between plants and also travel with the plants (da Graça *et al.*, 2016; ANSES, 2019; EFSA, 2019 b). In Florida (USA), infected samples of *D. citri* were found on oranges in fruit trailers, the insects were distributed throughout the loads on the fruit itself and not on accompanying plant debris (Halbert *et al.*, 2010).

PEST SIGNIFICANCE

Economic impact

Huanglongbing is currently regarded as one of the most important socio-economic threats to commercial citrus production at global level. Control measures in the field are difficult because long term resistance is still unknown and chemical-cultural management is technically difficult and expensive. In areas where the disease is endemic or uncontrolled, its progression within orchards can be relatively rapid. It is reported that it can reach more than 95 % of prevalence within 3 to 13 years after the onset of symptoms (Gottwald, 2010). Severe symptoms are always observed 5 to 8 years after planting in areas where abundant populations of bacteriferous vectors are prevailing. In diseased orchards, yield is reduced and fruit quality is affected. Yield reduction can reach 30 to 100 %, depending on the proportion of affected canopy and may render affected orchards non-economical within seven to ten years after planting. In many countries of America, Asia, and Africa, yield losses and difficulties in maintaining economically viable orchards have forced many growers out of business (Gottwald, 2010; Rasowo *et al.*, 2019).

In Asia, where ‘*Ca. Liberibacter asiaticus*’ occurs, huge impacts have been reported. A review about the disease in China (Zheng *et al.*, 2018) shows its enormous consequences for the citrus industry for over a hundred years. In Indonesia, 3 million trees were destroyed between 1960 and 1970 (Tirtawadja, 1980) and 4 million between 1986-1988 (Aubert, 1993). In India and Thailand, the disease was described as widespread and causing catastrophic losses during the 1960s and 1970s (Bové *et al.*, 1993; Varma *et al.*, 1993). In Saudi Arabia, all sweet oranges and mandarin trees had declined by 1986 leaving only limes (Aubert, 1993). In the Philippines, mandarin production decreased by

85 % in only eight years. In northern Bali, almost 100 % of mandarin trees planted in 1990-91 were severely affected five years later. In most cases, when the diseased trees were replaced, the disease reappeared on the newly planted trees. Almost 100 million trees have been affected and destroyed in many countries of South and Southeast Asia, compromising the local citriculture and large areas of citrus cultivation had to be abandoned (Gottwald, 2010).

In the Americas, the economic impact has also been dramatic, and the current situation is still difficult, despite the fact that the first detections were made in the 21st century and the causal agent was well known. In Brazil, five years after the identification of '*Ca. Liberibacter asiaticus*' in 2004 (Coletha-Filho *et al.*, 2004), more than 4 million trees were eliminated (officially and unofficially by the growers directly) in attempts to limit the dissemination of '*Ca. Liberibacter asiaticus*' and '*Ca. Liberibacter americanus*'. One year later, the number of symptomatic trees was estimated to be ca. 2 million (ca. 87 %) according to Belasque *et al.* (2010). In Florida (USA), costs of cultivation drastically increased since 2005, when the disease was first reported (Spren *et al.*, 2014). The costs of visual inspections of trees increased from 4 to 17 USD/ha and costs of insecticide treatments increased from 240 to 1000 USD/ha per year (Belasque *et al.*, 2010). The disease also had a very high economic and social impact during the first seven years after its detection. In addition, many packing houses and processing plants closed, with significant declines in employment and it was estimated that losses reached more than 3.63 billion USD in Florida and that more than 6 600 jobs were lost (Hodges & Spren, 2012).

The African form, '*Ca. Liberibacter africanus*', is considered to be less aggressive than the Asian form. However, da Graça & Korsten (2004) based on past information, reported that 4 out of 11 million trees in South Africa were affected by this disease. Crop losses of 30-100 % had been reported in South Africa during the 20th century and many of these orchards had subsequently to be abandoned or removed (Buitendag, 1991). In East Africa, surveys in Kenya and Tanzania, showed that it had the greatest impact on citrus production in the cooler highland regions, causing yield losses of 25–100 % (Rasowo *et al.*, 2019).

Finally, the economic impact of the disease caused by '*Ca. Liberibacter americanus*' in Brazil is difficult to determine due to the lack of data, and the fact that it currently appears to be displaced by the Asian form (CABI, 2018).

Control

Control options of huanglongbing have been evaluated for over a hundred years but as the disease situation and dynamics vary among countries, these options have been adapted. Consequently, it should be stressed that there is no universal solution for huanglongbing, but some strategies have been found to be useful in different areas.

In Africa, control of '*Ca. Liberibacter africanus*' during the second part of the 20th century relied on a combination of measures that were considered the most appropriate for each country (including chemical or biological vector control, trunk injections with tetracycline, pruning, thermotherapy, eradication, use of disease-free planting material, alternative hosts) and this integrated approach obtained relative success (da Graça, 1991; da Graça & Korsten, 2004). In South Africa, where most of the research was performed, removal of infected branches or trees, use of *Liberibacter*-free planting material, and control of the psyllid vector were applied (Buitendag & von Broembsen, 1993) with the main emphasis on the effect of systemic insecticides against *T. erytrae*, to maintain low psyllid populations.

In Asia, most research on strategies for a successful control of '*Ca. Liberibacter asiaticus*' were carried out in China with a similar approach, by promoting large-scale production of healthy nursery plants, early removal of infected plants in existing orchards, and applying insecticide sprays at critical flushing periods (Ke & Fan, 1990; Zheng *et al.*, 2018). In the Americas, the most comprehensive example for the management of '*Ca. Liberibacter asiaticus*' is provided by the Sao Paulo state in Brazil because it is one of the few regions in the world where control against huanglongbing has been carried out on a large scale by the growers, and found to be successful. It is summarized by Belasque *et al.* (2010) and based on three principles: (i) inoculum reduction by exhaustive inspections and frequent removal of affected trees, (ii) control of psyllid vector populations by insecticide treatments, to prevent new trees from becoming infected, and (iii) replanting with healthy tested plants produced under screen facilities only. Data from farms where the recommended measures have been applied since 2004 showed that the disease can be controlled. But the success was mainly obtained in large farms and it was necessary to apply the recommended measures rigorously. They must be strictly utilised in all the orchards of the area to be efficient. However, in a review on the management practices in Florida (antibiotics, insecticide applications, enhanced foliar nutritional programs, thermotherapy, and biological control), after the analysis of their economic performance and the economic

impact of several control options, Li *et al.* (2020) concluded that broad-spectrum insecticides provide the only cost-effective strategy for mitigating the high impact of the disease in the conditions of this state.

An essential part of the integrated control of huanglongbing in all the continents is the use of healthy plant material for replanting after eradication, developing a certification program based on microshoot-tip grafting *in vitro* (Navarro & Juárez, 2007; FAO, 2014), and producing plants in protected greenhouses to avoid the presence of vectors.

The successful management reported in some countries was considered a short-term solution to keep the citrus industry alive while other long-term solutions can be developed for an effective and integrated control of the disease.

Phytosanitary risk

In the EPPO region, host plants of huanglongbing are essentially *Citrus* species that are intensively cultivated in the Mediterranean basin countries. For the moment, none of the bacteria associated with huanglongbing has been found in the EPPO region, but the two vectors, *D. citri* and *T. erytrae*, are already present in some countries. There is no suggestion that native Mediterranean vectors could exist. If the huanglongbing associated bacteria were introduced in citrus-growing areas of the EPPO region, it is foreseen that tree development, harvest amount and quality would be severely impacted, and that this would ultimately seriously limit the citrus industry. Based on the experience of citrus-producing countries in other parts of the world, many changes quickly take place for the industry when new outbreaks are detected. In the short term, costly eradication, vector control, and nursery certification programs have to be immediately put in place and quarantine restrictions for export probably will appear. Nursery production must be maintained free of the disease, combined with the increased demand due to increasing infected tree removals. This can result in a rapid reduction of citrus production area as diseased trees are continuously removed (Gottwald, 2010) and final consequences are not only economic but also social and environmental. Considering the severity of huanglongbing, it is essential to keep this disease (and its vectors when possible) out of the EPPO region and to prevent their spread in the Middle East.

PHYTOSANITARY MEASURES

Considering the severity of huanglongbing, EPPO has recommended to prohibit the importation of citrus plants for planting and cut branches or buds of citrus from areas or countries where citrus huanglongbing (or either of its vectors) are present. In the EU, in addition, in areas where *T. erytrae* or *D. citri* occurs, control is compulsory, and it is prohibited to move plant material from infested areas to pest-free areas. In the EU territory, it is also forbidden to import fruit from third countries with their peduncles and leaves. In disease free countries as those of the Mediterranean area, awareness, monitoring, surveillance, pest risk assessment, quarantine measures and action plans are advised (Duran-Vila *et al.*, 2014; Siverio *et al.*, 2017). Procedures for official control with the aim of detecting, containing and eradicating huanglongbing and its vectors are provided in the EPPO Standard PM 9/27 (EPPO, 2020). As surveys should be carried out in all the EU member countries, a pest survey card was prepared by the European Food Safety Authority (EFSA, 2019) to assist EU Member States in planning their huanglongbing annual survey activities.

Healthy plant material is essential, and it should be available in the different EPPO countries. It can be obtained from citrus plants grown under quarantine restricted facilities, by using microshoot-tip grafting to produce pathogen-free buds *in vitro*. This pathogen-free material should be kept and propagated under insect-proof screenhouses, and its health status checked periodically, preferably by molecular techniques (e.g. real-time PCR) before being released or grafted onto indicator plants. Such a certification scheme is routinely used in Spain and in many countries where citrus are economically important crops (Navarro & Juárez, 2007).

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