

# EPPO Datasheet: *Clavibacter sepedonicus*

Last updated: 2021-11-22

## IDENTITY

**Preferred name:** *Clavibacter sepedonicus*

**Authority:** (Spieckermann & Kotthoff) Li et al.

**Taxonomic position:** Bacteria: Actinobacteria: Micrococcales: Microbacteriaceae

**Other scientific names:** *Bacterium sepedonicum* Spieckermann & Kotthoff, *Clavibacter michiganensis* subsp. *sepedonicus* (Spieckermann & Kotthoff) Davis et al., *Corynebacterium michiganense* subsp. *sepedonicum* (Spieckermann & Kotthoff) Carlson & Vidaver, *Corynebacterium sepedonicum* (Spieckermann & Kotthoff) Skaptason & Burkholder

**Common names:** bacterial ring rot of potato, ring rot of potato, vascular wilt of potato

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**EPPO Categorization:** A2 list

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**EU Categorization:** Quarantine pest ((EU) 2019/2072 Annex II B)

**EPPO Code:** CORBSE



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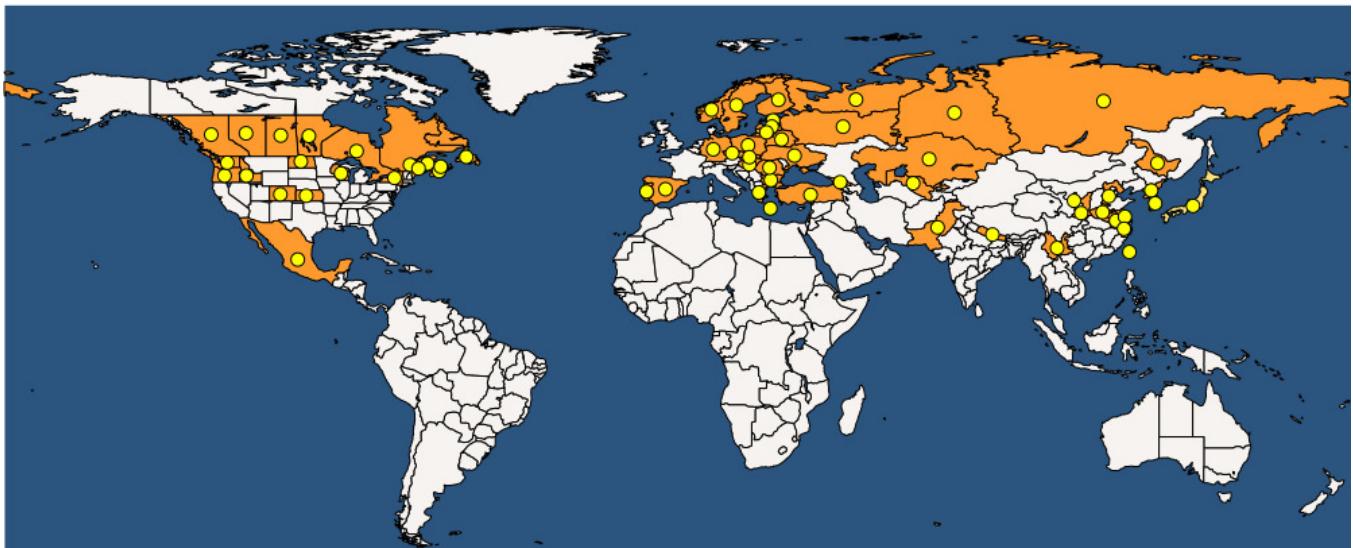
## HOSTS

Potato (*Solanum tuberosum*) is the only known major and significant host of *Clavibacter sepedonicus*. The bacterium has been isolated from symptomatic and asymptomatic sugar beet (*Beta vulgaris*). However, these findings are rare and appear to depend on the sugar beet variety (Bugbee *et al.*, 1987; Ignatov *et al.*, 2018; Van der Wolf *et al.*, 2005a). *C. sepedonicus* has also been isolated once from naturally infected tomato plants (Van Vaerenbergh *et al.*, 2016). Upon artificial inoculation, many members of the Solanaceae family (e.g. *Solanum melongena*), but also other plant species (e.g. *Urtica dioica*), were found to be susceptible to *C. sepedonicus* (Knorr, 1948; Van der Wolf *et al.*, 2005a).

**Host list:** *Beta vulgaris*, *Solanum lycopersicum*, *Solanum tuberosum*

## GEOGRAPHICAL DISTRIBUTION

*C. sepedonicus* was first described in Northern Europe and used to be found mainly in regions with a temperate climate in the northern hemisphere. Within the EPPO region, the climate in North, North-West and Central Europe is favorable to the disease. In the Southern part of the EPPO region, climatic conditions are not suitable for the establishment of ring rot except, in mountainous areas (Li *et al.*, 2018). In the EPPO region, *C. sepedonicus* is often reported with restricted distribution, and is only considered widespread in Russia, Ukraine and on the island of Crete (Greece).



## Clavibacter sepedonicus (CORBSE)

● Present

● Transient

2026-02-11

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**EPPO Region:** Belarus, Bulgaria, Czechia, Estonia, Finland, Georgia, Germany, Greece (mainland, Kriti), Hungary, Kazakhstan, Latvia, Lithuania, Norway, Poland, Portugal (mainland, Mainland Portugal), Romania, Russian Federation (Central Russia, Eastern Siberia, Northern Russia, Western Siberia), Slovakia, Spain (mainland), Sweden, Türkiye, Ukraine, Uzbekistan

**Asia:** China (Anhui, Hebei, Heilongjiang, Henan, Jiangsu, Ningxia, Shaanxi, Yunnan, Zhejiang), Japan, Kazakhstan, Korea, Democratic People's Republic of, Korea, Republic of, Nepal, Pakistan, Taiwan, Uzbekistan

**North America:** Canada (Alberta, British Columbia, Manitoba, New Brunswick, Newfoundland, Nova Scotia, Ontario, Prince Edward Island, Québec, Saskatchewan), Mexico, United States of America (Colorado, Idaho, Kansas, Maine, New York, North Dakota, Oregon, Washington, Wisconsin)

## BIOLOGY

The most common pathway of introduction of *C. sepedonicus* is through infected seed potatoes. True potato seeds might also be a source of infection if they come in contact with contaminated tools (Van der Gaag *et al.*, 2015). After a diseased seed potato is planted, the bacteria multiply very rapidly and pass along the vascular strands into the stems and petioles. From there they reach the roots and maturing daughter tubers, sometimes within 8 weeks after planting. The daughter tubers may themselves be used as seed potatoes and perpetuate the disease. *C. sepedonicus* does generally not survive in the soil during winter. The bacterium can, however, survive and remain infectious on potato bags, barn walls, machinery and other equipment and in volunteer plants from an infected crop. Survival is longest in cold dry conditions. *C. sepedonicus* is relatively vulnerable to higher temperatures ( $>55^{\circ}\text{C}$ ) suggesting that compost may not be a major inoculum source. However, survival of the bacterium inside protective plant tissue during the composting process might occur and the use of residues from potato processing in agriculture is therefore not recommended (Steinmöller *et al.*, 2013; Stevens *et al.*, 2021). The bacterium remains infectious at and above freezing temperatures for at least 18 months on burlap and for 63 months in infected potato stems (Nelson, 1985). If volunteer plants from a previously infected crop are lifted with an otherwise healthy seed potato crop, that crop can be infected. *C. sepedonicus* has a relatively low optimal growth temperature ( $21\text{-}23^{\circ}\text{C}$ ) and therefore it is mainly confined to cooler areas of the world (see Geographical distribution).

## DETECTION AND IDENTIFICATION

### Symptoms

The symptoms shown by infected plants are rather variable and, because they usually appear late in the growing season, may be mistaken for late blight (*Phytophthora infestans*), *Verticillium* wilt (*Verticillium albo-atrum*, *Verticillium dahliae*)

), stem canker (*Rhizoctonia solani*) or those caused by drought. The first symptoms of wilting develop in lower leaves, either all around the plant or only on one side of one stem. The margins of the leaves roll inwards and upwards and the surface loses its light shiny appearance. Leaves become progressively dull light-green, then grey-green with occasional mottling, then yellow and finally brown and necrotic. When infected stems are cut across, discolouration of vascular tissue is not obvious. Symptom formation is enhanced by hot, dry weather conditions (De Boer & Slack, 1984; Whitworth *et al.*, 2019).

Tuber symptoms may be confused with those caused by the bacterium *Ralstonia solanacearum* (EFSA, 2019). Tuber infection occurs through the stolon. Early infections can be observed, when the tuber is cut across the heel end, as narrow glassy to cream-yellow zones along the vascular tissue near the stolon end. In the case of more advanced infections this narrow yellowish to light-brown zone surrounds all the vascular tissue. In later stages the vascular ring and the discoloured zone become soft. Characteristically, upon squeezing, the tissue outside the vascular ring is easily separated from the inner tissues and creamy, cheese-like ribbons of odourless bacterial ooze with macerated tissue are expelled. In these advanced stages, external symptoms may also be observed, consisting of reddish to brown blotches around the eyes. The skin shows irregular, often star-shaped cracks. These cracked tubers are very susceptible to secondary soft-rot micro-organisms which obscure the ring rot symptoms (De Boer & Slack, 1984; Van der Wolf *et al.*, 2005b; Whitworth *et al.*, 2019). Mild infections in both susceptible and tolerant potato cultivars may cause so-called latent infections of daughter tubers. Latent infections can only be traced by laboratory testing (see Detection and inspection methods).

## Morphology

*C. sepedonicus* is a non-spore forming, non-motile, Gram-positive rod-shaped bacterium that forms white mucoid colonies (Hayward & Waterston, 1964; Li *et al.*, 2018).

## Detection and inspection methods

Surveillance for the presence of *C. sepedonicus* in a country or area not known to have potato ring rot, is usually based on a systematic detection survey. Specific guidance on the sampling of potato tubers in store or in the field (shortly before harvest) is given in the EPPO Standards PM 9/2 National regulatory control systems for *C. sepedonicus* (EPPO, 2011), PM 3/70 *Export certification and import compliance checking for potato tubers* (EPPO, 2019a) and PM 3/71 *General crop inspection procedure for potatoes* (EPPO, 2007). Additionally, samples may be inspected visually by cutting tubers at the stolon end, and growing potato crops may be visually inspected at appropriate times for typical signs and symptoms of the disease. It should be considered that under European climatic conditions, above ground symptoms are rarely found and then often only at the end of the season (EPPO, 2011).

Because symptoms of ring rot are variable and sometimes masked by other diseases, and because *C. sepedonicus* often is present without causing symptoms, ring rot can be confirmed only by laboratory testing. *C. sepedonicus* is a slow-growing bacterium and therefore when isolating the bacterium an enrichment step is often necessary to prevent it being overgrown by other bacteria. This can be done by inoculating specific eggplant varieties with potato extracts so the bacterium can multiply inside this plant. Subsequent isolation and purification steps are strongly facilitated by this step (EFSA, 2019; EPPO, 2022; Van der Wolf *et al.*, 2005b).

An immunofluorescence test and several molecular tests have been widely implemented in diagnostic laboratories to detect *C. sepedonicus*. Due to specificity problems observed in some cases, it is important to use a second test for detection, based on a different biological principle or on a different part of the genome, to confirm a positive result in the first detection test (EFSA, 2019; EPPO, 2022; Van der Wolf *et al.*, 2005b). The conventional PCR based test by Pastrik *et al.* (2000) and several real-time PCR tests have been shown to perform well in recent test performance studies (Vaerenbergh *et al.*, 2017; Vreeburg *et al.*, 2018). Among the real time PCR tests the one of Schaad *et al.* (1999) as well as the more recently developed real time PCR tests (Gudmestad *et al.*, 2009; Massart *et al.*, 2014; Vreeburg *et al.*, 2016; Vreeburg *et al.*, 2018), exhibit high analytical sensitivity and analytical specificity and have been implemented in diagnostic laboratories.

An updated version of the EPPO diagnostic protocol for the bacterium, providing details on the detection and identification tests is available (EPPO, 2022).

## PATHWAYS FOR MOVEMENT

Important means of spread are the planting of infected seed potatoes and contamination of containers, equipment and premises. When seed potatoes are cut before planting the cutting knife is an important dispersal unit: after cutting an infected tuber, 20-30 healthy tubers may be infected. Planters and graders which have been contaminated by bacteria from a few highly infected potatoes are also a potent infection source. Spread in the field from plant to plant is usually very low, but there is experimental evidence that some insects, including the Colorado beetle (*Leptinotarsa decemlineata*), leafhoppers and aphids can transmit the disease (Christie *et al.*, 1991; Duncan & Généreux, 1960; Mansfeld-Giese, 1997).

## PEST SIGNIFICANCE

### Economic impact

Damage is caused by destruction of vascular tissues and subsequent wilting and dying of plants and secondary rotting of tubers. In the past crop losses have been mainly reported from North America (up to 50%; Easton, 1979) and Russia (15-30% of plants infected, up to 47% crop loss; Muller & Ficke, 1974). Where ring rot occurs in the EPPO region, the disease appears more sporadically and at low levels of infection. However, a single infected tuber can already have a large economic impact. The economic impact can be caused by direct crop losses, by rejection of infected lots and by loss of (potential) export markets (Van der Wolf *et al.*, 2005b).

### Control

At the moment there is no method of direct chemical or biological control available. Breeding for resistance produced in the past some tolerant cultivars, which are not used much (Manzer *et al.*, 1987; Manzer & McKenzie, 1988). The most important methods of control are production of disease-free seed potatoes following strict certification and testing schemes (Nelson, 1985; EPPO, 1999) and sanitation (Lynch *et al.*, 1989).

In addition, crop rotation and weed/volunteer control are important preventive measures (EFSA, 2019; EPPO, 2020). Since the bacterium might be present in mixed soil from potato handling facilities, soil should only be returned to agricultural fields if the risk is considered acceptable. Conditions for returning soil to a place of production used to grow potatoes are described in draft Standard PM 3/92 (1) *Management of phytosanitary risks for potato crops resulting from movement of soil associated with root crops and potatoes* (EPPO, in press). Since the bacterium might survive inside protective plant tissue during the composting process, the use of residues from potato processing in agriculture is not recommended (Steinmöller *et al.*, 2013; Stevens *et al.*, 2021).

Disinfection is not part of routine hygiene measures but is obligatory after *C. sepedonicus* has been detected. EPPO developed a Standard that describes the cleaning and disinfection procedures in the potato production chain (EPPO, 2006). The efficacy of several chemical disinfection methods on different surfaces has been investigated (Howard *et al.*, 2015). More specifically, disinfection of wooden potato crates with a product containing sodium-*p*-toluenesulfochloramide has been shown to be effective (Stevens *et al.*, 2017).

### Phytosanitary risk

A number of seed-potato-producing countries in the EPPO region are free from the pest, as well as most Mediterranean countries exporting ware potatoes towards Northern European countries. The pathogen is likely to be able to establish wherever climatic conditions are favorable for the pathogen and potatoes are grown and to become increasingly widespread. While the direct economic impact of ring rot may only be moderate, especially with modern production systems, it would constitute a major extra constraint on seed potato production in countries where it does not occur, with considerable indirect effects on trade.

## PHYTOSANITARY MEASURES

Ring rot can occur at low levels in potato production systems and can cause latent infection of tubers. Therefore, phytosanitary measures focusing on potato consignments only are inadequate. Measures have to be implemented for the whole production system, i.e. on the material from which potato consignments are derived and at the place/site or area of production. For seed potatoes, in particular, they involve a series of multiple checks, each of which is considered by itself insufficient.

EPPO recommends that countries where *C. sepedonicus* is not known to occur, or which have implemented eradication or containment measures according to PM 9/2 (EPPO, 2011), should require measures for import of seed potatoes (except microplants and minitubers) and ware potatoes. According to EPPO Standard PM 8/1 (EPPO, 2017) seed and ware potatoes imported from a country where the pest occurs should be subject to transitional arrangements. Imported potatoes should come from a pest-free area and originate from a pest-free potato production and distribution system, according to EPPO Standard PM 3/61 (EPPO, 2019b), or the exporting country should have implemented an official regulatory control system according to EPPO Standard PM 9/2 (EPPO, 2011). If potatoes are imported from a country where *C. sepedonicus* is not known to occur, the absence should be confirmed by a survey following ISPM 6 *Surveillance* (IPPC, 2018). In addition, post-entry quarantine programs are established to allow safe movement of potato germplasm for research and breeding purposes (EPPO, 2019c).

## REFERENCES

Bugbee WM, Gudmestad NC, Secor GA & Nolte P (1987) Sugar beet as a symptomless host for *Corynebacterium sepedonicum*. *Phytopathology* **77**, 765-770.

Christie RD, Sumalde AC, Schulz JT & Gudmestad NC (1991) Insect transmission of the bacterial ring rot pathogen. *American Potato Journal* **68**, 363-372.

Davis MJ, Gillaspie AG, Vidaver AK & Harris RW (1984) *Clavibacter*: a new genus containing some phytopathogenic coryneform bacteria, including *Clavibacter xyli* subsp. *xyli* sp. nov., subsp. nov. and *Clavibacter xyli* subsp. *cynodontis* subsp. nov., pathogens that cause ratoon stunting disease of sugarcane and bermudagrass stunting disease. *International Journal of Systematic Bacteriology* **2**, 107-117.

De Boer SH & Slack SA (1984) Current status and prospects for detecting and controlling bacterial ring rot of potatoes in North America. *Plant Disease* **68**, 841-844.

Duncan J & Généreux H (1960) La transmission par les insectes de *Corynebacterium sepedonicum*. *Canadian Journal of Plant Science* **40**, 110-116.

Easton GD (1979) The biology and epidemiology of potato ring rot. *American Potato Journal* **56**, 459-460.

EFSA (2019) EFSA Panel on Plant Health: Bragard C, Dehnen-Schmutz K, Di Serio F, Gonthier P, Jaques Miret JA, Justesen AF, Macleod A, Magnusson CS, Milonas P, Navas-Cortes J, Parnell S, Potting R, Reignault PL, Thulke HH, Van Der Werf W, Vicent Civera A, Yuen J, Zappalà L, Van Der Wolf J, Kaluski T, Pautasso M & Jacques M. Pest categorisation of *Clavibacter sepedonicus*. *EFSA Journal* **17**, 1-26.

EPPO (1999) EPPO Standard PM 4/28(1) Seed potatoes. Certification schemes. *EPPO Bulletin* **29**, 253-267. Available at <https://gd.eppo.int/taxon/PSTVD0/documents>

EPPO (2022) Diagnostics. EPPO Standard PM 7/59 (2) *Clavibacter sepedonicus*. *EPPO Bulletin* **52**(2), 262-285. Available at <https://gd.eppo.int/standards/PM7/>

EPPO (2006) Phytosanitary treatments. EPPO Standard PM 10/1 (1) Disinfection procedures in potato production. *EPPO Bulletin* **36**, 463-466. Available at <https://gd.eppo.int/standards/PM10/>

EPPO (2007) Phytosanitary procedures. EPPO Standard PM 3/71 *General crop inspection procedure for potatoes*. *EPPO Bulletin* **37**, 592-597. Available at <https://gd.eppo.int/standards/PM3/>

EPPO (2011) National regulatory control systems. EPPO Standard PM 9/2 (2) *Clavibacter michiganensis* subsp. *sepedonicus*

. EPPO Bulletin **41**, 385-388. Available at <https://gd.eppo.int/standards/PM9/>

EPPO (2017) Commodity-specific phytosanitary measures. EPPO Standard PM 8/1(2) Potato. Commodity-specific phytosanitary measures. EPPO Bulletin **47**, 487-503. Available at <https://gd.eppo.int/taxon/CORBSE/documents>

EPPO (2019a) Phytosanitary procedures. EPPO Standard PM 3/70 *Export certification and import compliance checking for potato tubers*. EPPO Bulletin **36**, 423-424. Available at <https://gd.eppo.int/standards/PM3/>

EPPO (2019b) Phytosanitary procedures. EPPO Standard PM 3/61(2) Pest-free areas and pest-free production and distribution systems for quarantine pests of potato. EPPO Bulletin **49**, 480-481. Available at <https://gd.eppo.int/standards/PM3/>

EPPO (2019c) Phytosanitary procedures. EPPO Standard PM3/21(3) Post entry quarantine for potato. EPPO Bulletin **49**, 452-479. Available at <https://gd.eppo.int/standards/PM3/>

EPPO (2020) Phytosanitary procedures. EPPO Standard PM 3/89(1) Control of volunteer potato plants. EPPO Bulletin **50**, 372-382. Available at <https://gd.eppo.int/standards/PM3/>

EPPO (In press) Diagnostics. EPPO Standard PM 7/59(2) *Clavibacter sepedonicus*. EPPO Bulletin In press.

EPPO (In press) Phytosanitary procedures. EPPO Standard PM 3/92 (1) *Management of phytosanitary risks for potato crops resulting from movement of soil associated with root crops and potatoes*. EPPO Bulletin **51** In press.

Gudmestad NC, Mallik I, Pasche JS, Anderson NR & Kinzer K (2009) A Real-Time PCR assay for the detection of *Clavibacter michiganensis* subsp. *sepedonicus* based on the cellulase A gene sequence. *Plant Disease* **93**, 649-659.

Hayward AC & Waterston JM (1964) *Corynebacterium sepedonicum*. IMI Description of Fungi and Bacteria **2**, 14. Wallingford UK: CAB International.

Howard RJ, Harding MW, Daniels GC, Mobbs SL, Lisowski SLI & De Boer SH (2015) Efficacy of agricultural disinfectants on biofilms of the bacterial ring rot pathogen, *Clavibacter michiganensis* subsp. *sepedonicus*. *Canadian Journal of Plant Pathology* **37**, 273-284.

Ignatov AN, Panycheva JS, Spechenkova N & Taliantsky M (2018) First report of *Clavibacter michiganensis* subsp. *sepedonicus* infecting sugar beet in Russia. *Plant Disease* **102**, 2634.

IPPC (2018) ISPM 6 Surveillance. Rome, IPPC, FAO. Available at <https://www.ippc.int/en/core-activities/standards-setting/ispms/>

Knorr LC (1948) Suscept range of the potato ring rot bacterium. *American Potato Journal* **25**, 361-371.

Li X, Tambong J, Yuan K, Chen W, Xu H, Lévesque CA & De Boer SH (2018) Re-classification of *Clavibacter michiganensis* subspecies on the basis of whole-genome and multi-locus sequence analyses. *International Journal of Systematic and Evolutionary Microbiology* **68**, 234-240.

Lynch DR, Nelson GA & Kulcsar F (1989) Elimination of bacterial ring rot (*Corynebacterium sepedonicum* [Speck. and Kotth.] Skapt. and Burk.) by in vitro culture of sprout tissue. *Potato Research* **32**, 341-345.

Mansfeld-Giese K (1997) Plant-to-plant transmission of the bacterial ring rot pathogen *Clavibacter michiganensis* subsp. *sepedonicus*. *Potato Research* **40**, 229-235.

Manzer FE, Gudmestad NC & Nelson GA (1987) Factors affecting infection, disease development, and symptom expression of bacterial ring rot. *American Potato Journal* **64**, 671-676.

Manzer FE & McKenzie AR (1988) Cultivar response to bacterial ring rot infection in Maine. *American Potato Journal* **65**, 333-339.

Massart S, Nagy C & Jijakli MH (2014) Development of the simultaneous detection of *Ralstonia solanacearum* race

3 and *Clavibacter michiganensis* subsp. *sepedonicus* in potato tubers by a multiplex real-time PCR assay. *European Journal of Plant Pathology* **138**, 29-37.

Muller HJ & Ficke W (1974) [Bacterial ring rot (*Corynebacterium sepedonicum*) a dangerous quarantine disease for potato cultivation.]. *Nachrichtenblatt für den Pflanzenschutz in der DDR* **28**, 159-160 (in German).

Nelson GA (1985) Survival of *Corynebacterium sepedonicum* in potato stems and on surfaces held at freezing and above-freezing temperatures. *American Potato Journal* **62**, 23-28.

Nouiou I, Carro L, García-López M, Meier-Kolthoff JP, Woyke T, Kyrpides NC, Pukall R, Klenk H-P, Goodfellow M & Göker M (2018) Genome-based taxonomic classification of the phylum Actinobacteria. *Frontiers in Microbiology* **9**, 1-119.

Pastrik KH (2000) Detection of *Clavibacter michiganensis* subsp. *sepedonicus* in potato tubers by multiplex PCR with coamplification of host DNA. *European Journal of Plant Pathology* **106**, 155-165.

Schaad NW, Berthier-Schaad Y, Sechler A & Knorr D (1999) Detection of *Clavibacter michiganensis* subsp. *sepedonicus* in potato tubers by BIO-PCR and an automated real-time fluorescence detection system. *Plant Disease* **83**, 1095-1100.

Steinmöller S, Müller P, Bandte M & Büttner C (2013) Risk of dissemination of *Clavibacter michiganensis* ssp. *sepedonicus* with potato waste. *European Journal of Plant Pathology* **137**, 573-584.

Stevens LH, Lamers JG, Van Der Zouwen PS, Mendes O, Van Den Berg W, Tjou-Tam-Sin NNA, Jilesen CJTJ, Spoorenberg PM & Van Der Wolf JM (2017) Chemical eradication of the ring rot bacterium *Clavibacter michiganensis* subsp. *sepedonicus* on potato storage crates. *Potato Research* **60**, 145-158.

Stevens LH, Tom JY, van der Zouwen PS, Mendes O, Poleij LM & van der Wolf JM (2021) Effect of temperature treatments on the viability of *Clavibacter sepedonicus* in infected potato tissue. *Potato Research* **64**, 1-18.

Van Vaerenbergh J, Müller P, Elphinstone JG, Vreeburg RAM & Janse JD (2017) Euphresco inter?laboratory comparison (2009–2012) on detection of *Clavibacter michiganensis* subsp. *sepedonicus* and *Ralstonia solanacearum* in potato tubers: proposal to inc. *EPPO Bulletin* **47**, 24-32.

Van der Gaag (2015) Pest Risk Analysis EU internal movement of true potato seed (TPS) of registered TPS varieties: probability of association of regulated pests and analysis of risk reduction options. *Netherlands Food and Consumer Product Safety Authority*, 1-41. Available at <https://pra.eppo.int/getfile/40de6c9d-aa1d-4a0d-93f8-8b420ba1ddff>

Van der Wolf JM, Beckhoven JRCM, Hukkanen A, Karjalainen R & Muller P (2005a) Fate of *Clavibacter michiganensis* ssp. *sepedonicus*, the causal organism of bacterial ring rot of potato, in weeds and field crops. *Journal of Phytopathology* **153**, 358-365.

Van der Wolf JM, Elphinstone JG, Stead DE, Metzler M, Müller P, Hukkanen A & Karjalainen R (2005b) Epidemiology of *Clavibacter michiganensis* subsp. *sepedonicus* in relation to control of bacterial ring rot. *Plant Research International Report* **95**, 1-44.

Van Vaerenbergh J, De Paepe B, Hoedekie A, Van Malderghem C, Zaluga J, De Vos P & Maes M (2016) Natural infection of *Clavibacter michiganensis* subsp. *sepedonicus* in tomato (*Solanum tuberosum*). *New Disease Reports* **33**, 7.

Vreeburg RAM, Bergsma-Vlami M, Bollema RM, De Haan EG, Kooman-Gersmann M, Smits-Mastebroek L, Tameling WIL, Tjou-Tam-Sin NNA, Van De Vossenberg BTLH & Janse JD (2016) Performance of real-time PCR and immunofluorescence for the detection of *Clavibacter michiganensis* subsp. *sepedonicus* and *Ralstonia solanacearum* in potato tubers in routine testing. *EPPO Bulletin* **46**, 112-121.

Vreeburg RAM, Zendman AJW, Pol A, Verheij E, Nas M & Kooman-Gersmann M (2018) Validation of four real-time TaqMan PCRs for the detection of *Ralstonia solanacearum* and/or *Ralstonia pseudosolanacearum* and/or *Clavibacter michiganensis*

subsp. *sepedonicus* in potato tubers using a statistical regression approach. *EPPO Bulletin* **48**, 86-96.

Whitworth JL, Selstedt RA, Westra AAG, Nolte P, Duellman K, Yellareddygari SKR & Gudmestad NC (2019) Symptom expression of mainstream and specialty potato cultivars to bacterial ring rot (*Clavibacter sepedonicus*) and evaluation of in-field detection. *American Journal of Potato Research* **96**, 427-444.

### **EFSA resources used when preparing this datasheet**

EFSA Pest survey card on *Clavibacter michiganensis* subsp. *sepedonicus* <https://doi.org/10.2903/sp.efsa.2019.EN-1569>.

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### **How to cite this datasheet?**

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### **Datasheet history**

This datasheet was first published in the EPPO Bulletin in 1978, revised in the two editions of 'Quarantine Pests for Europe' in 1992 and 1997, as well as in 2021. It is now maintained in an electronic format in the EPPO Global Database. The sections on 'Identity', 'Hosts', and 'Geographical distribution' are automatically updated from the database. For other sections, the date of last revision is indicated on the right.

CABI/EPPO (1992/1997) *Quarantine Pests for Europe (1<sup>st</sup> and 2<sup>nd</sup> edition)*. CABI, Wallingford (GB).

EPPO (1978) EPPO Data Sheet on Quarantine Organisms no 51: *Corynebacterium sepedonicum*. *EPPO Bulletin* **8** (2), 25-29. <https://doi.org/10.1111/j.1365-2338.1978.tb02765.x>



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