**EPPO Datasheet: *Bactrocera occipitalis***

Last updated: 2020-09-23

**IDENTITY**

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| **Preferred name:** *Bactrocera occipitalis***Authority:** (Bezzi)**Taxonomic position:** Animalia: Arthropoda: Hexapoda: Insecta: Diptera: Tephritidae**Other scientific names:** *Chaetodacus ferrugineus occipitalis* (Bezzi), *Dacus occipitalis* (Bezzi)[view more common names online...](https://gd.eppo.int/taxon/BCTROC/)**EPPO Categorization:** A1 list[view more categorizations online...](https://gd.eppo.int/taxon/BCTROC/categorization)**EPPO Code:** BCTROC |  |

**Notes on taxonomy and nomenclature**

*Bactrocera occipitalis*belongs to the *B. dorsalis*species complex (see Drew & Hancock, 1994).

**HOSTS**

Known from a limited but varied list of hosts belonging to six different plant families. The USDA Compendium of Fruit Fly Host Information (CoFFHI) (Liquido *et al.,* 2019). provides an extensive host list with detailed references.

**Host list:** *Mangifera indica*, *Psidium guajava*, *fruit trees*

**GEOGRAPHICAL DISTRIBUTION**

*Bactrocera occipitalis*has a restricted distribution and is only recorded from islands in Maritime Southeast Asia, i.e. Borneo Island and the Philippine archipelago. Earlier records of *Bactrocera dorsalis* being introduced in Palau, appear to also comprise *B. occipitalis* (see [**Pacific Fruit Fly Project**](https://lrd.spc.int/species/bactrocera-occipitalis-bezzi)).

 **Asia:** Brunei Darussalam, Indonesia (Kalimantan), Malaysia (Sabah), Philippines

 **BIOLOGY**

Little is known about the biology of *B. occipitalis*. The general life cycle is considered similar to those of other *Bactrocera* species infesting fruits: eggs are deposited inside fruits by the female puncturing the fruit skin. Three larval stages develop inside the fruit, feeding on the plant tissue. Once mature the third instar larva will leave the fruit, dig down into the soil and turn into a pupa enclosed in a puparium. The adult fly will emerge from the puparium. No information is available regarding the duration of the life cycle or the environmental requirements.

**DETECTION AND IDENTIFICATION**

**Symptoms**

Attacked fruit have tiny oviposition punctures, but these and other symptoms of damage are often difficult to detect in the early stages of infestation. Considerable damage may occur inside the fruit before symptoms are visible externally, often as networks of tunnels accompanied by rotting.

**Morphology**

***Larva***

Fruit fly larvae in general have a typical shape, i.e., cylindrical maggot-shape, elongate, anterior end narrowed and somewhat recurved ventrally, with anterior mouth hooks, and flattened caudal end. Their length varies from 5 to 15 mm. Identification to species level is not possible based on larvae. A key for the 3rd-instar larvae is available in White & Elson- Harris (1992) and is useful for an identification to the genus level. The larvae of *B. occipitalis*have not been described in detail.

***Adult*** (after diagnostic description given by Drew & Romig, 2013. Additional character states character states of the female after Drew & Hancock, 1994)

Male
Face fulvous with a pair of large oval black spots; postpronotal lobes and notopleura yellow; scutum black except dark red-brown along posterior margin and enclosing prescutellar. setae, below and behind lateral postsutural vittae, around notopleural suture, around anterior margin of notopleura and inside postpronotal lobes; broad parallel-sided or subparallel lateral postsutural yellow vittae ending at intra-alar seta (in some specimens the vittae end behind the intra-alar seta); medial postsutural yellow vitta absent; mesopleural stripe reaching midway between anterior margin of notopleuron and anterior notopleural seta dorsally; scutellum yellow; legs with femora entirely fulvous; fore tibiae pale fuscous to fuscous, mid tibiae pale fuscous to fuscous basally tending paler apically, hind tibiae fuscous; wing with cells bc and c colourless, microtrichia in outer corner of cell c only; a narrow fuscous costal band distinctly overlapping R2+3 and widening markedly across apex of wing; a narrow fuscous anal streak; supernumerary lobe of medium development; abdominal terga III-V with a narrow transverse black band across anterior margin of tergum III and expanding to cover lateral margins, dark fuscous to black rectangular markings anterolaterally of tergum IV which sometimes continue to cover posterolateral margin of this tergum, dark fuscous to black anterolateral corners on tergum V, a very broad medial longitudinal black band over all three terga, a pair of oval orange-brown shining spots on tergum V; abdominal sterna dark coloured.

Female
As for male in the general body colour patterns. Wing, supernumerary lobe weak; pecten absent from abdominal tergum III. Ovipositor basal segment orange-brown, dorsoventrally compressed and tapering posteriorly in dorsal view; ratio of length of oviscape to length of tergum V, 0.68:1; aculeus apex needle shaped.

Remark: differentiation between this species and closely related species within the *B. dorsalis*species complex is difficult and needs expert confirmation. See ISPM 27 DP 29 (IPPC, 2019) for details on how to differentiate between the main species of commercial importance belonging to the species complex.

**DNA barcoding**

The molecular identification of *B. occipitalis* through DNA barcoding (COI) proves to be problematic as this species cannot be properly resolved from a number of closely related species, including species from the *B. dorsalis*species complex (see ISPM 27 DP 29 - IPPC, 2019). Additionally, the presence of unidentified / possibly misidentified reference sequence in the Barcoding Index Number Systems (BINs) in which this species is represented, might also bias its molecular ID. Sequences are available in the Barcode of Life Data Systems([**BOLD**](https://www.boldsystems.org/index.php/TaxBrowser_TaxonPage?taxid=79331)).

**Detection and inspection methods**

Males are attracted to methyl eugenol. Both sexes can be monitored by traps baited with protein-based attractants. Detection is also possible by examination of fruit for oviposition punctures and then rearing the larvae through to the adult stage.

**PATHWAYS FOR MOVEMENT**

Transport of infested fruits is the main mean of movement and dispersal to previously uninfested areas. Adult flight can also result in dispersal but previous citations of long (50-100 km) dispersal movements for *Bactrocera*spp. are unsubstantiated according to a recent review by Hicks *et al.* (2019). Dispersal up to 2 km is considered more typical.

**PEST SIGNIFICANCE**

**Economic impact**

The full impact of *B. occipitalis* is not fully understood because of co-occurrence with other similar species such as *B. dorsalis*. Although it is reported from a limited number of commercial fruits, no figures are available on the magnitude of infestation.

**Control**

Management for this species includes the general control measures for *Bactrocera* spp. (see Vargas *et al.* 2015 for an overview of management options). These include sanitation (to gather all fallen and infested host fruits and destroy them). Insecticidal protection is possible by using a cover spray or a bait spray. Bait sprays work on the principle that both male and female tephritids are strongly attracted to a protein source from which ammonia emanates. Bait sprays have the advantage over cover sprays in that they can be applied as a spot treatment so that the flies are attracted to the insecticide and there is minimal impact on natural enemies and other beneficials.

**Phytosanitary risk**

*Bactrocera occipitalis*is a known pest of peach in the area where it is present. It can be moved in trade with infested fruit. No detailed study has been made on climatic suitability of the EPPO region for this species, and it is unclear whether it could become established in the EPPO region. However, even transient populations could impact export of host fruit from the EPPO region. The EFSA Panel on Plant Health, in their Pest Categorization of non-EU Tephritidae (EFSA, 2020) placed *B. occipitalis* on the list of fruit flies that satisfy the criteria to be regarded as a potential Union quarantine pest for the EU.

**PHYTOSANITARY MEASURES**

Consignments of fruits from countries or regions where *B. occipitalis* occurs should be inspected for symptoms of infestation and those suspected should be cut open in order to look for larvae. Possible measures include that such fruits should come from an area where *B. occipitalis*does not occur, or from a place of production found free from the pest by regular inspection for 3 months before harvest. Plants transported with roots from countries or regions where *B. occipitalis* occurs should be free from soil, or the soil should be treated against puparia. The plants should not carry fruits.

**REFERENCES**

EFSA PLH Panel (EFSA Panel on Plant Health), Bragard C, Dehnen-Schmutz K, Di Serio F, Gonthier P, Jacques MA, Jaques Miret JA, Justesen AF, Magnusson CS, Milonas P, Navas-Cortes JA, Parnell S, Potting R, Reignault PL, Thulke HH, Van der Werf W, Vicent Civera A, Yuen J, Zappalà L, Bali EM, Papadopoulos N, Papanastassiou S, Czwienczek E & MacLeod A (2020) Pest categorization of non-EU Tephritidae. EFSA Journal 18: 5931, 62pp. <https://doi.org/10.2903/j.efsa.2020.5931>

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**CABI resources used when preparing this datasheet**

CABI Datasheet on Pest <https://www.cabi.org/isc/datasheet/8729>

**ACKNOWLEDGEMENTS**

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**How to cite this datasheet?**

EPPO (2024) *Bactrocera occipitalis*. EPPO datasheets on pests recommended for regulation. Available online. <https://gd.eppo.int>

**Datasheet history**

This datasheet was first published in 1997 in the second edition of 'Quarantine Pests for Europe', as part of the *Bactrocera* *dorsalis*species complex, and revised in 2020. It is now maintained in an electronic format in the EPPO Global Database. The sections on 'Identity', ‘Hosts’, and 'Geographical distribution' are automatically updated from the database. For other sections, the date of last revision is indicated on the right.

CABI/EPPO (1997) *Quarantine Pests for Europe (2nd edition).* CABI, Wallingford (GB).

