



ORGANISATION EUROPEENNE
ET MEDITERRANEENNE
POUR LA PROTECTION DES PLANTES

EUROPEAN AND MEDITERRANEAN
PLANT PROTECTION
ORGANIZATION

EPPO

Reporting

Service

Paris, 2002-08-01

Reporting Service 2002, No. 8

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2002/122 First reports of *Anastrepha obliqua* in Barbados and Grenada

The IPPC Secretariat recently informed the EPPO Secretariat of the introductions of *Anastrepha obliqua* (Diptera: Tephritidae – EPPO A1 quarantine pest) in Barbados and Grenada. The pest was found in November 2001 in Barbados, and in February 2002 in Grenada. *A. obliqua* was observed on *Psidium guajava* and *Spondias purpurea*. The situation of *A. obliqua* in both Barbados and Grenada can be described as follows: **Present, no details.**

Source: **FAO, IPPC Secretariat, 2002-07.**

Additional key words: new records

Computer codes: ANSTOB, BB, GD

2002/123 Situation of *Bactrocera zonata* in Réunion

In the Mascarene islands, *Bactrocera zonata* (Diptera, Tephritidae – EPPO A1 quarantine pest) was first detected in 1987 in Mauritius where it rapidly replaced other fruit flies. This triggered a surveillance programme in Réunion. *B. zonata* was first identified there in 1991, although it seems likely that it had been confused for some time with the indigenous *B. montyana*. Since 2000, a significant increase of *B. zonata* populations has been noticed in the north of the Island. To control the pest, the island was divided into 2 zones: a northern part where most captures were made and an eradication programme was initiated, and a southern part where captures remained rare and a surveillance programme was set up. In the eradication zone, methyl-eugenol traps (including an insecticide) were placed along a 1 x 1 km grid and checked every week. Male annihilation technique was used and ‘killer-blocks’ containing methyl-eugenol and malathion were placed (4-10 blocks/ha, renewed every 3 months). Hot-spot treatments were also made (protein bait + malathion) around the sites where captures had been made. In addition, host fruits (*Mangifera indica*, *Prunus persica*, *Psidium guajava*, *Terminalia catappa*) showing punctures were collected to check the presence of *B. zonata*. In the south part of the island, more than 100 traps were placed in orchards, near the main fruit and vegetable markets, and near the seaport. When males were captured, trapping was intensified and, if new findings were made, control actions (use of killer-blocks) were undertaken. After two years of these intensive measures, it was unfortunately observed that *B. zonata* had established near St Denis (although at low populations), and continued to progress outside the eradication zone into the surveillance zone where it was found in numerous places. In May 2001, the eradication campaign was abandoned in the north of the island. At the end of 2001, the highest populations were found in the north-east (St Denis, St Marie, St André), north-west (La Possession, Rivière des Galets, St Paul) and in the west. Observations made on host fruits showed that *B. zonata* could multiply on: *Citrus paradisi*, *Citrus reticulata*, *Mangifera indica*, *Mimusops elengi*, *Persea americana*, *Psidium guajava*, *Punica granatum*, *Terminalia catappa*, *Syzygium*



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jambos, *Ziziphus jujuba*. Nevertheless, intensive control will continue to be applied in Réunion to contain the pest. Studies on host plants, ecology and biology of *B. zonata*, as well as on efficacy of chemical treatments will be carried out, and more information will be provided to the growers. The situation of *B. zonata* in Réunion can be described as follows: **Present, found mainly on the north and west of the island, under official control.**

Source: Hurtrel, B.; Quilici, S.; Jeuffrault, E.; Manikom, R.; Georger, S.; Gourdon, F. (2002) Etat de siège contre la mouche de la pêche, *Bactrocera zonata*. Bilan des opérations de lutte menées à la Réunion.
Phytoma – La Défense des Végétaux n° 551, 18-21.

Additional key words: eradication, detailed record

Computer codes: DACUZO, RE

2002/124 PRAs on *Ceratitis capitata* and *C. rosa* in Martinique

In the French Antilles, only one species of fruit fly is present *Anastrepha obliqua* (Diptera, Tephritidae - EPPO A1 quarantine pest), and causes minor problems. However, many other fruit fly species could be introduced via the intensive and diverse trade of fresh fruits and vegetables from other parts of the world. A network of traps has been set up in Martinique for 13 years. Various types of traps (for *Anastrepha*, *Bactrocera* and *Ceratitis* species) are installed near waste dumps, sea ports, airport and orchards. Traps are inspected every week, and catches are identified. For the moment, only *A. obliqua* has been found but an action plan is ready in case of other findings. In addition to these practical field activities, two Pest Risk Analyses (PRAs) have recently been conducted in Martinique for *Ceratitis capitata* (EPPO A2 quarantine pest) and *Ceratitis rosa* (EPPO A1 quarantine pest), following the EPPO PRA scheme. The conclusion was that *C. capitata* presented a high potential for introduction (large imports of host fruits, tourists introducing many host fruits and plants illegally) and a high potential for establishment (favourable climate, great diversity of host plants). Economic losses may be limited in commercial orchards, but costs of treatments are likely to be substantial, and the main risk is for the environment with the colonization of many host plants. For *C. rosa*, it is estimated that the risk of introduction is lower, as only imports of litchis and citrus fruits coming from South Africa by plane, or tourists carrying these fruits illegally, are likely to introduce the pest. However, the potential for establishment is similar to *C. capitata*. As a consequence of these PRAs, the fruit fly trapping network was intensified in Martinique.

Source: Landau, E.; Bertrand, P.; Davidas, M.A.; Guéret, C. (2002) Les mouches des fruits menacent-elles la Martinique ? Réseau de surveillance et analyses de risque phytosanitaire vis-à-vis de *Ceratitis capitata* et *Ceratitis rosa*.
Phytoma – La Défense des Végétaux, n° 551, 22-25.

Additional key words: PRA

Computer codes: CERTCA, CERTRO, MT



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2002/125 New records of *Liriomyza trifolii* in South America and the Caribbean

A paper from Martinez & Etienne (2002) gives a list of 456 species of Agromyzidae present in the neotropical region with details on their geographical distribution. From this list, the EPPO Secretariat has noted the following new records for *Liriomyza trifolii* (EPPO A2 quarantine pest):

Argentina
Chile (Juan Fernandez Islands)
Mexico
Saint Martin (Netherlands Antilles/Guadeloupe)
Virgin islands (not specifying US or British)

Source: Martinez, M.; Etienne, J. (2002) Liste systématique et biogéographique des Agromyzidae (Diptera) de la région néotropicale.
Bollettino di Zoologia Agraria e di Bachicoltura, Serie II, 34(1), 25-52.

Additional key words: new records

Computer codes: LIRITR

2002/126 First report of *Impatiens necrotic spot tospovirus* in Iran

In Iran, *Impatiens necrotic spot tospovirus* (EPPO A2 quarantine pest) was detected for the first time in commercial nurseries and field-grown ornamentals in Mahallat and Tehran provinces. The virus was detected in *Rosa*, *Gazania*, *Dendranthema*, *Leucanthemum*, *Matricaria camomilla*, *Pelargonium roseum*, *Salvia*, *Dianthus caryophyllus* in the province of Mahallat, and on *Gazania* and *Bougainvillea spectabilis* in the Tehran province. A few samples (*Dendranthema* and *Leucanthemum*) were infected by both *Tomato spotted wilt* and *Impatiens necrotic spot tospovirus*. The situation of *Impatiens necrotic spot tospovirus* in Iran can be described as follows: **Present, found on ornamentals in Mahallat and Tehran provinces.**

Source: Shahraneen, N.; Ghotbi, T.; Mahraban, A.H.; (2002) Occurrence of *Impatiens necrotic spot virus* in ornamentals in Mahallat and Tehran Provinces in Iran.
Plant Disease, 86(6), p 694.

Additional key words: new record

Computer codes: INSV00, IR



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2002/127 First report of *Tomato infectious chlorosis crinivirus* in Spain

In Spain, during summer and autumn 2001, symptoms of yellowing, bronzing, brittleness and rolling of lower leaves were observed in protected and field-grown tomatoes in the Castellón province, Cataluña. The presence of *Tomato infectious chlorosis crinivirus* (EPPO Alert List) was detected in diseased tomatoes. This is the first report of this virus in Spain. The situation of *Tomato infectious chlorosis crinivirus* in Spain can be described as follows: **Present, found in protected and field-grown tomatoes in Cataluña.**

Source: Font, I.; Martínez-Culebras, P.; Jorda, M.C., Louro, D.; Vaira, A.M.; Accotto, G.P. (2002) First report of *Tomato infectious chlorosis virus* in Spain. **Plant Disease, 86(6), p 696.**

Additional key words: new record

Computer codes: TICV00, ES

2002/128 PCR method to differentiate between Tomato yellow leaf curl Sardinia and Tomato yellow leaf curl begomoviruses

Tomato yellow leaf curl-Sardinia and Tomato yellow leaf curl-Israel begomoviruses (TYLCV-Sar and TYLCV, both EPPO A2 quarantine pests) cannot be differentiated on the basis of their symptomatology, although TYLCV-Is is more aggressive and causes greater economic losses. A rapid PCR method has been developed in Spain to differentiate between the two virus species. A combination of primers (specific to any isolate of TYLCV and of TYLCV-Sar) was selected to develop a duplex PCR method which can detect either TYLCV-Sar or TYLCV.

Source: Martínez-Culebras, P.V.; Font, I.; Jordá, C. (2001) A rapid PCR method to discriminate between Tomato yellow leaf curl virus isolates. **Annals of applied Biology, 139(2), 251-257.**

Additional key words: diagnostics

Computer codes: TYLCV0



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2002/129 Distribution of TYLCV-Sar and TYLCV around the Mediterranean Basin

In the European and Mediterranean region two species of tomato yellow leaf curl viruses are present: TYLCV-Sar (first reported in 1989) and TYLCV (first reported in 1996). Around the Mediterranean Basin, their respective geographical distributions are as follows:

- **TYLCV-Sar**

Morocco, Italy (Sicilia, Sardegna), Spain (south of mainland, Islas Canarias)

- **TYLCV**

Israel, Morocco, Portugal (Algarve only), Spain (south of mainland, Islas Canarias), Tunisia, Turkey

Source: Moriones E. & Accotto, D.P (2000) Tomato yellow leaf curl virus complex (TYLCV). **EWSN Identification Guide Sheet, release no. 1, part B, p 1.**

Additional key words: detailed records

Computer codes: TYLCV, ES,IL, IT, MA, PT, TN, TR

2002/130 Phytosanitary measures against *Monilinia fructicola* in France

After the discovery of *Monilinia fructicola* (EPPO A1 quarantine pest) in southern France (see EPPO RS 2002/003), phytosanitary measures were decided. A compulsory control programme is applied over the whole country and concerns host plants of *M. fructicola* (almond, apricot, cherry, peach, plum). In commercial orchards: prophylactic action is taken to reduce infection as much as possible, fungicide treatments are applied following the advice given by warning services, after harvest all remaining fruits are destroyed, and harvesting machinery is disinfested. In packing stations: a general disinfection is done at the beginning of the season, all rejected fruits are destroyed as soon as possible. In orchards producing scions (certified or not): production of fruits is limited (strict thinning), after harvest all remaining fruits are eliminated, scions should be free from cankers. If scions are taken from other places, this should be reported to the official authorities a month in advance, and authorization is given only if conditions required for commercial orchards are met and symptoms absent. In nurseries producing young plants: fungicide treatments are applied at the beginning of the season and in autumn (leaf fall).

Source: Adminet – Journal Officiel (France). Arrêté du 3 avril 2002 relatif à la lutte contre *Monilia fructicola*, champignon parasite en vergers.
<http://www.admi.net/jo/20020419/AGRG0200772A.html>

Additional key words: phytosanitary measures

Computer codes: MONIFC, FR



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2002/131 Situation of *Xanthomonas axonopodis* pv. *dieffenbachiae* in Réunion

Xanthomonas axonopodis pv. *dieffenbachiae* (EPPO A1 quarantine pest) was first observed in Réunion in October 1997 in one glasshouse producing *Anthurium*, and the identity of the bacterium was confirmed in November 1997 (see EPPO RS 2001/040). Investigations showed that the source of infection was the import of contaminated plants from the Netherlands. All infected plants were destroyed. However, despite prophylactic measures taken on the production site concerned, at the end of 1998, the bacterium had spread to the whole site (3 ha of glasshouses). Today, production has not started again on this site. During 1997/1998, 3 other outbreaks were discovered, whose origin was again the import of contaminated lots from the Netherlands. As this clearly showed that the disease was threatening all growers on the island, strict measures were taken. Only imports of *in vitro* plants from nurseries found free from the disease by appropriate testing are now allowed. Importers must be registered, as well as the sites where plants are kept in quarantine for 18 months (until first flowering). Eradication is carried out in all production sites found infected: plants are destroyed (burning and burial in quicklime). Finally, information is provided to growers. In 2001, all infested plants were destroyed and on these previously infected sites no host plants were grown. Most recent surveys done on the main production sites showed that there are still 2 outbreaks which are now under eradication. In addition, the bacterium was detected in a few private gardens. Research is being carried out in Réunion to collect and characterize isolates of the bacterium from the island and other countries, and to develop a specific PCR method. The situation of *X. axonopodis* pv. *dieffenbachiae* in Réunion can be described as follows: **Present, found at 2 production sites and in a few private gardens, under eradication.**

Source: Jeuffrault, E.; Soustrade, I.; Laurent, P.; Gagnevin, L.; Henry, G. (2002) Dépérissement de l'anthurium dû à *Xanthomonas axonopodis* pv. *dieffenbachiae*. Bilan de trois ans de lutte à la Réunion : vigilance !
Phytoma – La Défense des Végétaux n° 551, 46-48.

Additional key words: detailed record

Computer codes: XANTDF, RE



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2002/132 *Ralstonia solanacearum* on *Anthurium* in Martinique

Martinique is a producer of *Anthurium* cut flowers, but intensification of this crop in the 1980s has led to the appearance of phytosanitary problems, in particular caused by bacteria. At present, 3 bacteria have been identified on *Anthurium* in Martinique: *Xanthomonas axonopodis* pv. *dieffenbachiae* (EPPO A1 quarantine pest) in 1984 (its finding led to the destruction of all infested plots), *Acidovorax anthurii* in 1986 (it rarely kills plants but flower quality is drastically reduced), and finally *Ralstonia solanacearum* (EPPO A2 quarantine pest) in 1999. Symptoms were characterized by water-soaked lesions along the midrib, large black necrotic lesions on leaves surrounded by a water-soaked margin on the underside, water-soaked lesions on flowers, yellowing bacterial ooze when cutting stem bases. Affected plants finally die. From 1999 to 2001, surveys were carried out on *Anthurium* and other host plants. Results showed a slow progression of the disease in *Anthurium* crops, mainly in the areas of Gros Morne, Saint Joseph and Morne Rouge. *R. solanacearum* was not found on other host plants (Araceae, Strelitziaceae, Zingiberaceae), with the exception of one sample of *Heliconia* (Heliconiaceae) which showed wilt symptoms. *Anthurium* isolates were characterized and found closely related to those causing Moko disease of banana (absent in Martinique). Pathogenicity tests were immediately conducted on banana, but no symptoms were obtained on artificial inoculation. In addition, during surveys carried out in banana orchards located near *Anthurium* infected plots, the bacterium was not found in banana. *Anthurium* isolates are different from Solanaceous isolates usually found in the Caribbean. In addition, similar types of isolates were discovered in December 2001 on Cucurbitaceae (cucumber, melon, *Cucurbita moschata*) showing symptoms of bacterial wilt. Further studies will be carried out on this new group of isolates attacking *Anthurium* and Cucurbitaceae in Martinique.

Source: Mian, D.; Coranson-Beaudu, R.; Duféal, D.; Grassart, L.; Mention, P. (2002) *Ralstonia solanacearum* sur anthurium à la Martinique. Une bactériose inquiétante.

Phytoma – La Défense des Végétaux, n° 551, 43-45.

Additional key words: new host plants

Computer codes: PSDMSO, MT



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2002/133 Details on the taxonomy and biology of *Rhizoecus hibisci*

The root mealybug *Rhizoecus hibisci* (Homoptera: Pseudococcidae – EPPO A1 quarantine pest) is frequently intercepted by the Netherlands on bonsais from China. In general, root mealybugs cause slow plant growth. Serious damage was only noticed once on imported *Serissa foetida* in a glasshouse in the Netherlands in 1992. Usually, damage is rather limited, despite the presence of high numbers of *R. hibisci*. In general, little information is available on the biology and morphology of pre-adult stages of this pest. These aspects were studied in the laboratory in the Netherlands. It was observed that at 21°C, one generation lasted 61 days on *Serissa* and much longer on *Nerium* (approximately 90 days). Numbers of eggs in ovisacs varied from 11 to 84 on different hosts. On average, eggs hatch after 9 days. 3 nymphal stages were observed followed by pupa and adult stages. Although males were never seen in intercepted bonsais, they could be observed in the laboratory on *Serissa*. A list of host plants gathered from the literature and observations is given: *Crinum asiaticum* (Amaryllidaceae), *Nerium oleander* (Apocynaceae), *Dieffenbachia* (Araceae), *Cryptanthus* (Bromeliaceae), *Dichorisandra thyrsiflora* (Commelinaceae), *Carex* (Cyperaceae), *Pelargonium* (Geraniaceae), *Hakonechloa macra* (Poaceae), *Cuphea hyssopifolia* (Lythraceae), *Hibiscus rosa-sinensis* (Malvaceae), *Calathea makoyama* (Marantaceae), *Ficus benjamina* (Moraceae), *Areca*, *Kentia*, *Phoenix canariensis*, *P. roebelenii*, *Rhapis excelsa*, *Sabal* (Arecaceae), *Sageretia thea* (Rhamnaceae), *Serissa foetida* (Rubiaceae), *Celtis*, *Zelkova serrata* (Ulmaceae). Finally, detailed descriptions of nymphal instars are also presented.

Source: Jansen, M.G.M. (2001) Instar identification and some notes about the life cycle of *Rhizoecus hibisci* Kawai & Takagi (Coccoidea: Pseudococcidae). **Bollettino di Zoologia Agraria e di Bachicoltura, Serie II, 33(3), 53-56.**

Additional key words: biology, host plants, taxonomy

Computer codes: RHIOHI



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2002/134 Situation of *Aonidiella citrina* in Italy

As reported in EPPO RS 95/035, a small outbreak of *Aonidiella citrina* (Homoptera: Diaspididae – EU Annexes) was found in 1994, in Calabria, Italy. Surveys and observations were made in citrus groves in 1995-1996 and in 1999-2000, in Calabria and Sicilia. 10 years after its first report, *A. citrina* is still confined to a limited area of the Sibari Plain in Calabria. It was not found in Sicilia. Biological observations showed that, in Calabria, *A. citrina* completes 3 generations per year, with peaks of crawlers in the first half of June, beginning of August and beginning of September. During winter, various stages can be observed (mainly the second instar). In March, mortality reached 65% due to abiotic and biotic factors, and in particular due to the activity of *Aphytis melinus* and *Encarsia citrina*. The situation of *Aonidiella citrina* in Italy can be described as follows: **Present, found in a limited area of the Sibari Plain in Calabria.**

Source: Longo, S.; Mazzeo, V.; Palmeri, D.; Benfatto, D.; Maurello, S.; di Leo, A. (2001) New remarks on the distribution and biology of *Aonidiella citrina* (Coquillet) (Homoptera: Coccoidea) in Italy.
Bollettino di Zoologia Agraria e di Bachicoltura, Serie II, 33(3), 508-509.

Additional key words: detailed record

Computer codes: AONDCI, IT

2002/135 Introduction of *Ceroplastes ceriferus* into Italy: addition to the EPPO Alert List

In January 2001, during a survey on scale populations of ornamental plants in parks and gardens near Verona (Veneto, Italy), an unusual species was noticed (Mori *et al.*, 2001). It was identified as *Ceroplastes ceriferus* (Homoptera, Coccidae), a polyphagous species thought to originate from Asia. This is the first report of this species in Europe. In Italy, it was collected from: *Acer palmatum*, *Cornus sanguinea*, *C. alba*, *Desmodium penduliflorum*, *Deutzia gracilis*, *Laurus nobilis*, *Magnolia stellata*, *Malus domestica*, *Spiraea japonica*, *Viburnum dentatum*. So far, *C. ceriferus* has been recorded outdoors in gardens and nurseries in two regions of northern Italy: Veneto (district of Verona) and Lombardia (district of Bergamo). The observation of overwintering adult females suggested that the pest is probably already established. No damage to plants has been observed, probably because of its recent introduction. Considering the high fecundity of *C. ceriferus*, its wide host-range and its already known ability to overwinter outdoors, the authors concluded that in absence of any control, this new pest is likely to spread and represent a threat for several ornamental species.



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Ceroplastes ceriferus (Homoptera, Coccidae) – Japanese or Indian wax scale

Why	<i>Ceroplastes ceriferus</i> came to our attention because of its recent introduction into Italy. It is a highly polyphagous pest which could represent a threat for ornamental plants, and possibly fruit crops.
Where	<p>It occurs in many parts of the world, but until recently it was still absent from Europe.</p> <p>Europe: Italy (reported for the first time in 2001 in Lombardia and Veneto on various ornamentals). <i>C. ceriferus</i> has been intercepted by the Netherlands in 1999 and 2000, on <i>Ficus</i> and <i>Podocarpus</i> from Taiwan.</p> <p>Asia: Cambodia, China, India, Indonesia, Japan, Malaysia, Myanmar, Philippines, Sri Lanka, Taiwan, Thailand, Vietnam.</p> <p>Africa: Malawi, Tanzania, Uganda.</p> <p>North America: Mexico, USA (present in many states)</p> <p>Central & South America, Caribbean: Brazil, Chile, Jamaica, Panama, Puerto Rico, US Virgin Islands.</p> <p>Oceania: Australia, Cook Islands, Fiji, Guam, New Caledonia, Papua New Guinea, New Zealand, Tonga, Vanuatu</p>
On which plants	Highly polyphagous (more than 122 plant species in 46 families). It attacks a wide range of crops, mostly fruit crops (e.g. apple, avocado, citrus, fig, pear, plum, quince, <i>Vaccinium</i> and many tropical fruit crops) and ornamentals (e.g. <i>Acer</i> , <i>Berberis</i> , <i>Buxus</i> , <i>Cornus</i> , <i>Deutzia</i> , <i>Euonymus</i> , <i>Ficus</i> , <i>Ilex</i> , <i>Lagerstroemia</i> , <i>Laurus</i> , <i>Magnolia</i> , <i>Platanus</i> , <i>Populus</i> , <i>Pyracantha</i> , <i>Rhododendron</i> , <i>Salix</i> , <i>Viburnum</i> ...)
Damage	<i>C. ceriferus</i> occurs on foliage, stems and branches. The scales can cause chlorotic spotting on the leaves which may fall prematurely, dieback of stems and wilting, and reduced vigour of the plants. Severe infestations disfigure plants because of the large numbers of white scales and copious honeydew on which sooty mould develops. <i>C. ceriferus</i> has one generation per year and overwinters as mature females. Mature females are covered with thick white wax, usually with an anteriorly projecting horn of wax. Immature stages (crawlers) are flattened and tiny. In North America, <i>C. ceriferus</i> is considered as a serious pest of ornamentals. It is also reported as a pest of tea in China (Guizhou), of poplars in India (Karnataka) and as a minor pest of avocado in Australia (Queensland).
Dissemination	Immature stages can move over short distances. Over long distances, all stages can be transported on infected plant material.
Pathway	Plants for planting, cut branches and foliage of host plants from countries where <i>C. ceriferus</i> occurs.
Possible risks	<i>C. ceriferus</i> is a highly polyphagous pest, and many of its host plants are grown in Europe for fruit production or ornamental purposes. Its recent introduction into Italy showed that the pest is able to survive in parts of Europe. Although more data is needed on the impact of this scale on fruit crops, it seems that it is more a threat for ornamental crops. Control of scales is usually difficult in practice, although biological control agents exist in other parts of the world, it is not known whether they could reduce populations sufficiently under European conditions.
Source(s)	<p>CABI Crop Protection Compendium 2001.</p> <p>Mori, N.; Pellizzari, G.; Tosi, L. (2001) <i>Ceroplastes ceriferus</i> (Fabricius) (Homoptera, Coccoidea): new pest of ornamentals in Europe? Bollettino di Zoologia Agraria e di Bachicoltura, Serie II, 33(3), 331-336.</p> <p>Mori, N.; Pellizzari, G.; Tosi, L. (2001) [First record of the wax scale <i>Ceroplastes ceriferus</i> (Fabricius) (Homoptera, Coccoidea) in Italy]. Informatore Fitopatologico, no. 10, 41-43.</p>
EPPO RS 2002/135 Panel review date	-
	Entry date 2002-08



EPPO *Reporting Service*

2002/136 Invasive plant species of concern in Denmark

The following species are listed as invasive plant species on the web site of the Danish Forest and Nature Agency:

Acer pseudoplatanus
Aegopodium podagraria
Ailanthus altissima
Elodea canadensis
Erigeron (Conyza) canadensis
Heracleum mantegazzianum
Impatiens parviflora
Impatiens glandulifera
Lupinus polyphyllus
Pastinaca sativa
Petasites hybridus
Pinus mugo
Pinus contorta
Prunus serotina
Rosa rugosa
Reynoutria japonica
Reynoutria sachalinensis
Robinia pseudoacacia
Solidago canadensis
Solidago gigantea
Spartina anglica
Spiraea douglasii
Spiraea ssp.
Telekia speciosa

Source: **Web site of the Danish Forest and Nature Agency:**
<http://www.sns.dk/natur/groen/a01.htm>

Additional key words: invasive alien species

Computer codes: DK



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2002/137 US draft Federal noxious weed lists

USDA-APHIS (US) has a noxious weed program which is designed to prevent the introduction into the United States of non-indigenous invasive plants and to prevent the spread of newly introduced invasive plants within the United States. On its web site, a draft list (2000-08-09) of noxious weeds is available. In a draft action plan for the noxious weeds program, this list has been further worked (2002-01), and plant species have been separated into four lists: federal noxious weeds, not in US (A1), federal noxious weeds, introduced (A2), regulated non-quarantine weeds (C) and a list of non-regulated weeds of interest to APHIS.

• **A1 Weeds: Federal noxious weeds, not in US**

<i>Aeginetia</i> spp. (Orobanchaceae)	Parasitic
<i>Alectra</i> spp. (Scrophulariaceae)	Parasitic
<i>Azolla pinnata</i> (Azollaceae)	mosquito fern, water velvet
<i>Carthamus oxyacantha</i> (Asteraceae)	wild safflower - Terrestrial
<i>Cuscuta</i> spp. other than native or introduced species (Cuscutaceae)	doddies - Parasitic
<i>Digitaria abyssinica</i> (= <i>D. scalarum</i>) (Poaceae)	African couch grass - Terrestrial
<i>Drymaria arenarioides</i> (Caryophyllaceae)	lightening weed, alfombrilla - Terrestrial
<i>Lagarosiphon major</i> (Hydrocharitaceae)	Oxygen weed - Aquatic/Wetland
<i>Leptochloa chinensis</i> (Poaceae)	Asian sprangletop - Terrestrial
<i>Lycium ferocissimum</i> (Solanaceae)	African boxthorn - Terrestrial
<i>Mikania cordata</i> (Asteraceae)	mile-a-minute - Terrestrial
<i>Monochoria hastata</i> (Pontederiaceae)	monochoria - Aquatic/Wetland
<i>Nassella trichotoma</i> (Poaceae)	serrated tussock - Terrestrial
<i>Opuntia aurantiaca</i> (Cactaceae)	jointed prickly pear - Terrestrial
<i>Orobanche</i> spp. other than native or introduced species (Orobanchaceae)	broomrapes - Parasitic
<i>Oryza longistaminata</i> (Poaceae)	red rice - Terrestrial
<i>Oryza punctata</i> (Poaceae)	red rice - Terrestrial
<i>Prosopis alpataco</i> (Fabaceae)	mesquites - Terrestrial
<i>Prosopis argentina</i> (Fabaceae)	mesquites - Terrestrial
<i>Prosopis articulata</i> (Fabaceae)	mesquites - Terrestrial
<i>Prosopis burkartii</i> (Fabaceae)	mesquites - Terrestrial
<i>Prosopis caldenia</i> (Fabaceae)	mesquites - Terrestrial
<i>Prosopis calingastana</i> (Fabaceae)	mesquites - Terrestrial
<i>Prosopis campestris</i> (Fabaceae)	mesquites - Terrestrial
<i>Prosopis castellanosi</i> (Fabaceae)	mesquites - Terrestrial
<i>Prosopis denudans</i> (Fabaceae)	mesquites - Terrestrial
<i>Prosopis elata</i> (Fabaceae)	mesquites - Terrestrial
<i>Prosopis ferox</i> (Fabaceae)	mesquites - Terrestrial
<i>Prosopis fiebrigii</i> (Fabaceae)	mesquites - Terrestrial
<i>Prosopis hassleri</i> (Fabaceae)	mesquites - Terrestrial
<i>Prosopis humilis</i> (Fabaceae)	mesquites - Terrestrial
<i>Prosopis kuntzei</i> (Fabaceae)	mesquites - Terrestrial
<i>Prosopis palmeri</i> (Fabaceae)	mesquites - Terrestrial



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A1 continued

<i>Prosopis rojasiana</i> (Fabaceae)	mesquites - Terrestrial
<i>Prosopis ruizlealii</i> (Fabaceae)	mesquites - Terrestrial
<i>Prosopis ruscifolia</i> (Fabaceae)	mesquites - Terrestrial
<i>Prosopis sericantha</i> (Fabaceae)	mesquites - Terrestrial
<i>Prosopis torquata</i> (Fabaceae)	mesquites - Terrestrial
<i>Rubus fruticosus</i> (Rosaceae)	wild blackberry complex - Terrestrial
<i>Rubus moluccanus</i> (Rosaceae)	wild blackberry - Terrestrial
<i>Sparganium erectum</i> (Sparganiaceae)	exotic bur-reed - Aquatic/Wetland
<i>Spermacoce alata</i> (Rubiaceae)	borreria - Terrestrial
<i>Striga</i> spp. other than native or introduced species (Scrophulariaceae)	witchweeds - Parasitic

• A2 Weeds: Federal noxious weeds introduced

<i>Ageratina adenophora</i> (Asteraceae)	crofton weed - Terrestrial
<i>Alternanthera sessilis</i> (Amaranthaceae)	sessile joyweed - Terrestrial
<i>Asphodelus fistulosus</i> (Liliaceae)	onionweed - Terrestrial
<i>Avena sterilis</i> L. (Poaceae)	animated or wild oat - Terrestrial
<i>Caulerpa taxifolia</i> (Caulerpaceae)	caulerpa - Aquatic/Wetland
<i>Chrysopogon aciculatus</i> (Poaceae)	pilipiliula - Terrestrial
<i>Commelina benghalensis</i> (Commelinaceae)	Benghal dayflower - Terrestrial
<i>Crupina vulgaris</i> (Asteraceae)	common crupina - Terrestrial
<i>Digitaria velutina</i> (Poaceae)	velvet fingergrass - Terrestrial
<i>Eichhornia azurea</i> (Pontederiaceae)	anchored waterhyacinth - Aquatic/Wetland
<i>Emex australis</i> (Polygonaceae)	three-cornered jack - Terrestrial
<i>Emex spinosa</i> (Polygonaceae)	devil's thorn - Terrestrial
<i>Galega officinalis</i> (Fabaceae)	goatsrue - Terrestrial
<i>Heracleum mantegazzianum</i> (Apiaceae)	giant hogweed - Terrestrial
<i>Hydrilla verticillata</i> (Hydrocharitaceae)	Hydrilla - Aquatic/Wetland
<i>Hypophila polysperma</i> (Acanthaceae)	Miramar weed - Aquatic/Wetland
<i>Imperata brasiliensis</i> (Poaceae)	Brazilian satintail - Terrestrial
<i>Imperata cylindrica</i> (Poaceae)	cogongrass - Terrestrial
<i>Ipomoea aquatica</i> (Convolvulaceae)	Chinese waterspinach - Aquatic/Wetland
<i>Ischaemum rugosum</i> (Poaceae)	murain-grass - Terrestrial
<i>Limnophila sessiliflora</i> (Scrophulariaceae)	ambulia - Aquatic/Wetland
<i>Melaleuca quinquenervia</i> (Myrtaceae)	melaleuca - Aquatic/Wetland
<i>Melastoma malabathricum</i> (Melastomataceae)	Terrestrial
<i>Mikania micrantha</i> (Asteraceae)	mile-a-minute - Terrestrial
<i>Mimosa diplotrichia</i> (Fabaceae)	giant sensitive plant - Terrestrial
<i>Mimosa pigra</i> (Fabaceae)	catclaw mimosa - Terrestrial
<i>Monochoria vaginalis</i> (Pontederiaceae)	pickerel weed - Aquatic/Wetland
<i>Orobanche minor</i> (Orobanchaceae)	Parasitic
<i>Orobanche ramosa</i> (Orobanchaceae)	Parasitic
<i>Oryza rufipogon</i> (Poaceae)	red rice - Terrestrial
<i>Ottelia alismoides</i> (Hydrocharitaceae)	duck-lettuce - Aquatic/Wetland
<i>Paspalum scrobiculatum</i> (Poaceae)	Kodo-millet - Terrestrial
<i>Pennisetum clandestinum</i> (Poaceae)	kikuyugrass - Terrestrial
<i>Pennisetum macrourum</i> (Poaceae)	African feathergrass - Terrestrial
<i>Pennisetum pedicellatum</i> (Poaceae)	kyasuma-grass - Terrestrial
<i>Pennisetum polystachion</i> (Poaceae)	missiongrass - Terrestrial



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A2 continued

<i>Prosopis farcta</i> (Fabaceae)	mesquites - Terrestrial
<i>Prosopis pallida</i> (Fabaceae)	mesquites - Terrestrial
<i>Prosopis reptans</i> (Fabaceae)	mesquites - Terrestrial
<i>Prosopis strombulifera</i> (Fabaceae)	mesquites - Terrestrial
<i>Rottboellia cochinchinensis</i> (Poaceae)	itchgrass - Terrestrial
<i>Saccharum spontaneum</i> (Poaceae)	wild sugarcane - Terrestrial
<i>Sagittaria sagittifolia</i> (Alismataceae)	arrowhead - Aquatic/Wetland
<i>Salsola vermiculata</i> (Chenopodiaceae)	wormleaf salsola - Terrestrial
<i>Salvinia auriculata</i> (Salviniaceae)	giant salvinia - Aquatic/Wetland
<i>Salvinia molesta</i> (Salviniaceae)	giant salvinia - Aquatic/Wetland
<i>Setaria pumila</i> (Poaceae)	cattail grass - Terrestrial
<i>Solanum tampicense</i> (Solanaceae)	wetland nightshade - Aquatic/Wetland
<i>Solanum torvum</i> (Solanaceae)	turkeyberry - Terrestrial
<i>Solanum viarum</i> (Solanaceae)	tropical soda apple - Terrestrial
<i>Striga asiatica</i> (Scrophulariaceae)	witchweed- Parasitic
<i>Striga gesnerioides</i> (Scrophulariaceae)	indigo witchweed - Parasitic
<i>Tridax procumbens</i> (Asteraceae)	coat buttons - Terrestrial
<i>Urochloa panicoides</i> (Poaceae)	liverseed grass - Terrestrial

• B. Regulated non-quarantine weeds

<i>Acroptilon repens</i> (Asteraceae)	Russian knapweed - Terrestrial
<i>Cardaria draba</i> (Brassicaceae)	hoary cress - Terrestrial
<i>Cardaria pubescens</i> (Brassicaceae)	white top - Terrestrial
<i>Cirsium arvense</i> (Asteraceae)	Canada thistle - Terrestrial
<i>Convolvulus arvensis</i> (Convolvulaceae)	field bindweed - Terrestrial
<i>Elytrigia repens</i> (Poaceae)	quack grass - Terrestrial
<i>Euphorbia esula</i> (Euphorbiaceae)	leafy spurge - Terrestrial
<i>Sonchus arvensis</i> (Asteraceae)	perennial sowthistle - Terrestrial
<i>Sorghum halepensis</i> (Poaceae)	Johnsongrass - Terrestrial

• C. Non-regulated weeds of interest to APHIS

<i>Alliaria petiolata</i> (Brassicaceae)	garlic mustard - Terrestrial
<i>Carduus nutans</i> (Asteraceae)	musk thistle - Terrestrial
<i>Centaurea diffusa</i> (Asteraceae)	diffuse knapweed - Terrestrial
<i>Centaurea maculosa</i> (Asteraceae)	spotted knapweed - Terrestrial
<i>Centaurea solstitialis</i> (Asteraceae)	yellow starthistle - Terrestrial
<i>Centaurea virgata</i> ssp. <i>squarrosa</i> (Asteraceae)	squarrose knapweed - Terrestrial
<i>Chondrilla juncea</i> (Asteraceae)	rush skeletonweed - Terrestrial
<i>Cynoglossum officinale</i> (Boraginaceae)	houndstongue - Terrestrial
<i>Cyperus esculentus</i> (Cyperaceae)	yellow nutsedge - Aquatic/Wetlands
<i>Cyperus rotundus</i> (Cyperaceae)	purple nutsedge - Aquatic/Wetlands
<i>Eichhornia crassipes</i> (Pontederiaceae)	Waterhyacinth - Aquatic/Wetlands
<i>Hieracium aurantiacum</i> (Asteraceae)	orange hawkweed - Terrestrial
<i>Hieracium pratense</i> (Asteraceae)	meadow hawkweed - Terrestrial
<i>Linaria dalmatica</i> (Scrophulariaceae)	Dalmatian toadflax - Terrestrial
<i>Linaria vulgaris</i> (Scrophulariaceae)	yellow toadflax - Terrestrial
<i>Lythrum salicaria</i> (Lythraceae)	purple loosestrife - Aquatic/Wetlands
<i>Myriophyllum spicatum</i> (Haloragaceae)	Eurasian watermilfoil - Aquatic/Wetlands



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Onopordum acanthium (Asteraceae)

Phragmites australis (Poaceae)

Polygonum perfoliatum (Polygonaceae)

Pueraria lobata (Fabaceae)

Rosa multiflora (Rosaceae)

Tamarix ramosissima (Tamaricaceae)

Scotch thistle - Terrestrial

common reed - Aquatic/Wetlands

mile-a-minute weed - Terrestrial

kudzu - Terrestrial

multiflora rose - Terrestrial

saltcedar - Terrestrial

Source: USDA-APHIS - US Federal noxious weed list.

<http://www.aphis.usda.gov/ppq/weeds>

Draft Action Plan for the Noxious Weeds Program

<http://www.aphis.usda.gov/ppq/weeds/weedsjan2002-pub.pdf>

Additional key words: invasive plants

Computer codes: US

2002/138 **New IPPC web site**

The IPPC Secretariat announced that its new web site (International Phytosanitary Portal) was launched on the 1st of August. The new address is:

http://www.ippc.int/cds_ippc/IPP/En/default.htm

Currently navigation is only in English (this will change rapidly over the next few weeks), but documents are available in other languages where appropriate.

Source: **IPPC Secretariat, 2002-08.**