

## Data Sheets on Quarantine Pests

**Potato Andean latent tymovirus****IDENTITY**

**Name:** Potato Andean latent tymovirus

**Synonyms:** Potato (Andean) latent virus Andean potato latent virus

**Taxonomic position:** Viruses: *Tymovirus*

**Common names:** APLV (acronym)

**Notes on taxonomy and nomenclature:** APLV is sometimes considered to be a strain of eggplant mosaic tymovirus but sequence comparisons show them to be distinct species.

**EPPO computer code:** POALXX

**EPPO A1 list:** No. 244

**EU Annex designation:** I/A1

**HOSTS**

The principal host of APLV is potatoes (*Solanum tuberosum*). The virus can also be transmitted mechanically to species of Amaranthaceae, Chenopodiaceae, Cucurbitaceae and Solanaceae (Gibbs *et al.*, 1966; Fribourg *et al.*, 1977).

**GEOGRAPHICAL DISTRIBUTION**

**EPPO region:** Absent.

**South America:** Widespread in the Andean countries Bolivia, Colombia, Ecuador and Peru, especially at high altitudes (Koenig *et al.*, 1979). Also recorded from Argentina, Chile and Paraguay (unconfirmed).

**EU:** Absent.

**BIOLOGY**

APLV belongs to the tymovirus group, members of which are typically beetle-transmitted. In experiments APLV was transmitted by a flea beetle (*Epitrix* sp.) with low efficiency. *Epitrix* spp. could act as natural vectors but only when high populations are present. The virus is readily transmitted by contact and at a low frequency through true potato seed (Jones & Fribourg, 1977). Transmission to tubers is erratic.

APLV was considered to be a strain of eggplant mosaic tymovirus (see Identity), but its isolates differ in several characteristics (Gibbs & Harrison, 1969, 1973; Angarita, 1977; Angarita & Dekegel, 1979). They can be combined into three major strain groups recognized by spur formation tests (Koenig *et al.*, 1979). Isolates of the Col-Caj-group and of the CCC-group have been found more commonly in the northern Andean region and the members of the Hu-group in the southern Andes from central Peru to Bolivia.

## **DETECTION AND IDENTIFICATION**

### **Symptoms**

The symptoms vary depending on virus strain, potato cultivar and growing conditions. They range from mild to severe mosaic with necrotic flecking, curling and leaf-tip necrosis. A wide daily fluctuation in temperature, in particular cold conditions, seems to favour symptom expression in infected plants growing at high altitude. Severe symptoms are also induced in mixed infections with other potato viruses (Jones & Fribourg, 1978).

### **Morphology**

The virus contains isometric particles of about 30 nm in diameter which sediment in two components: empty protein shells and nucleoprotein particles (Gibbs *et al.*, 1966).

### **Detection and inspection methods**

#### **Indicator plants**

*Nicotiana bigelovii* seems to be the best host for detecting APLV. Local lesions are developed in inoculated leaves followed by a systemic mosaic and characteristic systemic netting of minor veins. *Nicotiana clevelandii* shows necrotic or chlorotic spots on inoculated leaves with most isolates and a strong systemic mosaic with netting of the minor veins.

#### **Serological detection methods**

High-titre antisera can be prepared. The latex agglutination test detects all known isolates with the same antiserum, but it is less sensitive than ELISA (Koenig & Bode, 1977; Fribourg & Nakashima, 1984). ELISA is rather strain-specific (Koenig *et al.*, 1979), but this can be overcome by using a mixture of antisera for each of the three virus strain groups (Schroeder & Weidemann, 1990). Dot-ELISA on nitrocellulose membranes has been successful (CIP, 1989).

## **MEANS OF MOVEMENT AND DISPERSAL**

APLV is transmitted by contact between plants, insect vectors, possibly through tubers (transmission is erratic) and by true seed. In international trade, APLV could in principle be carried by potato tubers or by true seed of potato.

## **PEST SIGNIFICANCE**

### **Economic impact**

Despite its name, APLV can cause serious symptoms in secondarily infected potato plants (see Detection and identification); however, yield reductions have not been studied, and it is not clear how important APLV is in practice.

### **Control**

As with all potato viruses, control depends on the production of high-quality seed potatoes from virus-free nuclear stock.

### **Phytosanitary risk**

APLV is included among the non-European potato viruses of the EPPO A1 quarantine list (OEPP/EPPO, 1984a). It is also considered of quarantine concern by NAPPO and, in general, all regional plant protection organizations outside South America recommend very strict measures for potato material from that continent. The principal perceived risk is the introduction of new viruses into seed potato production schemes, increasing the cost and difficulty of operating these schemes, and opening up new possibilities for yield losses from single or mixed virus infections. Any seed potato-exporting country in which APLV

was reported would immediately find itself in difficulties with respect to the phytosanitary certification of its exports. The risk is particularly important because of the simple pathway which exists from useful germplasm material (local potato cultivars, wild tuber-forming *Solanum* spp.) in the potato's centre of diversity in South America through to nuclear stock material of new cultivars in seed potato-producing countries. Thus there is a great risk of introduction due to the increased international exchange of breeding material and germplasm, whether in the form of tubers, rooted cuttings, *in vitro* cultures or true seeds.

Individually, APLV could be regarded, among the group of South American potato pathogens, as of major importance for the EPPO region. It is distinguished, despite its name, by being a damaging disease, and it is also transmitted by true seed. Though it can relatively easily be excluded by prohibition of commercial trade in potato tubers, there is a risk of introduction with breeding material, in which it could only be detected by careful testing under quarantine.

## PHYTOSANITARY MEASURES

Importation of potato tubers from countries where APLV occurs should be prohibited. APLV is one of the group of South American pests of potato which justify strict post-entry quarantine procedures in the EPPO region, together with equivalent checks before export. Only material for scientific purposes, in quantities limited to what is strictly necessary and subject to import permit, should normally be imported from countries where APLV occurs. Because of the probability that any material of wild tuber-forming *Solanum* spp. originates ultimately from South America, the same tests should be applied whatever the origin. EPPO's specific quarantine requirements (OEPP/EPPO, 1990) outline suitable quarantine measures, while EPPO's phytosanitary procedures lay down the test procedures to be followed both before export and in post-entry quarantine after import (OEPP/EPPO, 1984b).

## BIBLIOGRAPHY

- Angarita, A. (1977) Characteristics of an Andean potato latent virus isolate from Colombia. *Parasitica* **33**, 103-110.
- Angarita, A.; Dekegel, D. (1979) Electrophoretic properties of a strain of Andean potato latent virus from Colombia and electron microscopy of virus-infected cells. *Parasitica* **35**, 16-24.
- CIP (1989) *Annual Report, International Potato Center (CIP), Lima, Peru*, p. 59.
- Fribourg, C.E.; Jones, R.A.C.; Koenig, R. (1977) Host plant reaction, physical properties and serology of three isolates of Andean potato latent virus from Peru. *Annals of Applied Biology* **86**, 373-380.
- Fribourg, C.E.; Nakashima, J. (1984) An improved latex agglutination test for routine detection of potato viruses. *Potato Research* **27**, 237-249.
- Gibbs, A.J.; Harrison, B.D. (1969) Eggplant mosaic virus, and its relationship to Andean potato latent virus. *Annals of Applied Biology* **64**, 225-231.
- Gibbs, A.J.; Harrison, B.D. (1973) Eggplant mosaic virus. *CMI/AAB Descriptions of Plant Viruses* No. 124. Association of Applied Biologists, Wellesbourne, UK.
- Gibbs, A.J.; Hecht-Poinar, E.; McKee, R.K. (1966) Some properties of three related viruses: Andean potato latent, dulcamara mottle and ononis yellow mosaic. *Journal of General Microbiology* **44**, 177-193.
- Jones, R.A.C.; Fribourg, C.E. (1977) Beetle, contact and potato true seed transmission of Andean potato latent virus. *Annals of Applied Biology* **86**, 123-128.
- Jones, R.A.C.; Fribourg, C.E. (1978) Symptoms induced by Andean potato latent virus in wild and cultivated potatoes. *Potato Research* **21**, 121-127.
- Koenig, R.; Bode, O. (1977) Sensitive detection of Andean potato latent and Andean potato mottle viruses in potato tubers with the serological latex test. *Phytopathologische Zeitschrift* **92**, 275-280.

- Koenig, R.; Fribourg, C.E.; Jones, R.A.C. (1979) Symptomatology, serological, and electrophoretic diversity of isolates of Andean potato latent virus from different regions of the Andes. *Phytopathology* **69**, 748-752.
- OEPP/EPPO (1984a) Data sheets on quarantine organisms No. 128, Potato viruses (non-European). *Bulletin OEPP/EPPO Bulletin* **14**, 11-22.
- OEPP/EPPO (1984b) Quarantine procedures No. 21, Potato viruses (non-European) and potato spindle tuber viroid. *Bulletin OEPP/EPPO Bulletin* **14**, 73-76.
- OEPP/EPPO (1990) Specific quarantine requirements. *EPPO Technical Documents* No. 1008.
- Schroeder, M.; Weidemann, H.L. (1990) Detection of quarantine viruses of potato by ELISA. *Bulletin OEPP/EPPO Bulletin* **20**, 581-590.